

Metabolic Health in the Transition Period and Fertility of Dairy Cows

R.S. Bisinotto, DVM, MS; E.S. Ribeiro, DVM, MS; F.S. Lima, DVM, MS; N. Martinez, DVM, MPVM; L.F. Greco, DVM, MS; J.E.P. Santos, DVM, MS, PhD

Department of Animal Sciences, University of Florida, Gainesville, FL 32611

Corresponding author: Dr. J.E.P. Santos, Email: jepsantos@ufl.edu

Abstract

During early postpartum, high-producing dairy cows undergo a period of extensive tissue catabolism because of negative nutrient balance. Homeorhetic controls assure that nutrients are partitioned to favor lactation at the same time that homeostasis secures survival. However, unrestrained metabolic disturbances often lead to diseases which, in turn, dramatically decrease both productive and reproductive performance. Negative nutrient balance has been associated with compromised immune and reproductive functions in dairy cows. Low circulating concentrations of glucose and insulin associated with elevated concentrations of non-esterified fatty acids and ketone bodies postpartum have detrimental effects on the function and viability of the oocyte, granulosa cells, and immune cells. Therefore, minimizing the extent and duration of negative nutrient balance in early lactation is expected to reduce morbidity and enhance fertility. Reductions in circulating concentrations of Ca, and vitamins A and E around parturition are also linked with impaired immune competence, and have to be accounted for when formulating peripartum diets. Furthermore, dietary additives that influence rumen or intermediary metabolism to favor postpartum health, and supplementation with specific fatty acids during early lactation and the breeding period, might benefit fertility of dairy cows. Finally, manipulating the length of the dry period and the caloric content of prepartum diets might have carry-over effects during the subsequent lactation that favor resumption of postpartum ovulation. Proactive management of dairy cows during the periparturient period is needed for cows to achieve high production with good fertility.

Résumé

Tôt en postpartum, les vaches laitières en forte production font face à une période de catabolisme important des tissus en raison d'un bilan nutritionnel négatif. Le contrôle homéorhétic assure que les nutriments soient bien partagés afin de favoriser la lactation au même moment où l'homéostasie maintient la survie. Toutefois, des déséquilibres métaboliques sans restrictions vont souvent mener à des maladies

qui peuvent à leur tour diminuer dramatiquement la performance de production et de reproduction. Un bilan nutritionnel négatif a été associé à la fragilisation des fonctions immunitaires et reproductives chez les vaches laitières. De faibles concentrations circulantes de glucose et d'insuline associées à des concentrations plus élevées d'acides gras non-estérifiés et de corps cétoniques en postpartum ont des effets adverses pour l'oocyte, la granulosa et les cellules immunitaires tout en nuisant à leur fonction et viabilité. Par conséquent, minimiser l'importance et la durée du bilan nutritionnel négatif en début de lactation devrait réduire la morbidité et accroître la fertilité. Une réduction des concentrations circulantes de Ca et des vitamines A et E au moment de la parturition a aussi été reliée à l'affaiblissement de la compétence immunitaire et devrait être considérée lors de la formulation du régime alimentaire en péripartum. De plus, les additifs alimentaires qui influencent le rumen ou le métabolisme intermédiaire pour favoriser la santé postpartum de même que l'apport supplémentaire d'acides gras particuliers tôt en lactation et durant la période de reproduction devraient favoriser la fertilité chez les vaches laitières. Finalement, la manipulation de la longueur de la période de tarissement et de l'apport calorique du régime prépartum pourrait avoir des répercussions lors de la lactation subséquente en favorisant le retour de l'ovulation postpartum. La régie proactive des vaches laitières en péripartum est nécessaire pour permettre aux vaches d'avoir une production plus élevée et une meilleure fertilité.

Introduction

Reproductive efficiency of the lactating herd is a major component of profitability in dairy farms. Reproduction determines when primiparous cows become multiparous leading to increments in milk yield; alters the average milk yield per day of calving interval; affects the number of replacement animals available and the risk of culling; and influences the rate of genetic progress. Unfortunately, improving fertility is not trivial. The establishment and maintenance of a pregnancy to term are affected by several genetic, physiological, and environmental factors that can be manipulated in order to sustain high fertility. Although causality is not always

established, it is well described that poor metabolic health negatively influences reproduction in dairy cows. The energetic status of a cow modulates the secretion of hormones that play key roles in growth of ovarian follicles, ovulation, corpus luteum (CL) formation, and oocyte competence. Furthermore, extensive lipolysis and products from fat metabolism may be detrimental to oocyte competence and subsequent embryo development. In addition, impaired metabolic health often leads to immunosuppression and the occurrence of diseases that further reduce fertility.

In order to maintain a 12- to 13-month calving interval, cows must become pregnant in the first few months postpartum, soon after periods of extensive tissue catabolism. Therefore, it is not a surprise that when negative nutrient balance is extended further in lactation, fertility is suppressed. Managing cows in the transition period in a proactive manner to minimize nutrient imbalances is mandatory for proper health, and it often benefits reproduction. The aim of this review is to discuss the potential mechanisms by which shifts in metabolism in early lactation influence fertility of dairy cows and propose nutritional strategies to improve metabolic health and enhance reproduction.

Prevalence of Diseases Postpartum and Impact on Fertility of Dairy Cows

The transition from the non-lactating pregnant to non-pregnant lactating state requires the high-producing dairy cow to drastically adjust its metabolism so that nutrients can be partitioned to support milk synthesis, a process referred to as homeorrhexis. A sharp increase in nutrient requirements generally occurs when feed intake is depressed in early lactation, which causes extensive mobilization of body tissues, particularly body fat, but also amino acids, minerals, and vitamins. Despite tight homeostatic controls and homeorrethic adjustments to cope with the changes in metabolism caused by milk production, 45 to 60% of dairy cows across different levels of milk production, breeds, and management systems develop metabolic and infectious diseases in the first months of lactation.^{58,61}

Calving-related disorders and diseases that affect the reproductive tract are major contributors to depression of fertility. Dystocia, metritis, and clinical endometritis were observed in 14.6, 16.1, and 20.8% of postpartum dairy cows in large US confinement herds, respectively.⁶¹ Cows that presented at least one of the aforementioned disorders were 50 to 63% less likely to resume ovarian cyclicity by the end of the voluntary waiting period, and 25 to 38% less likely to become pregnant following the first artificial insemination (AI) postpartum, compared with healthy cows. Moreover, cows with dystocia and those diagnosed with clinical endometritis were 67 and

55% more likely to lose their pregnancies during the first 60 days of gestation than healthy cows. The negative effects of reproductive disorders on subsequent fertility are also observed in dairy cows kept under grazing systems.⁵⁸ Even though the prevalence of dystocia, metritis, and clinical endometritis are numerically lower in grazing-based herds (8.2, 5.7, and 14.7%, respectively), cows with metritis had 2.7-fold increased odds of being anovular at 50 days postpartum compared with unaffected herd-mates. Cows affected with uterine diseases had marked depression in pregnancy at the first postpartum AI and increased risk of pregnancy loss. In fact, when diseases were classified as clinical (calving problem, metritis, clinical endometritis, mastitis, pneumonia, digestive problems, lameness), subclinical (subclinical hypocalcemia, subclinical ketosis, and severe negative energy balance based on excessive plasma nonesterified fatty acids), or both, affected cows had increased anovulation, reduced pregnancy per AI, and increased pregnancy loss⁵⁸ (Table 1). These data strongly suggest that diseases in early lactation have a profound impact on fertility of dairy cows, and maintaining metabolic health to minimize the risk of clinical and subclinical health problems is expected to benefit reproduction.

Negative Nutrient Balance Impacts Health and Reproduction of Dairy Cows

Increased nutrient needs associated with suppression of appetite generally drive dairy cows into a state of negative energy balance (NEB), which is often observed in the last week of gestation and the first two months postpartum. Under normal conditions, dry matter intake increases from 21.1 lb (9.6 kg) per day in the week preceding parturition to more than 48.4 lb (22 kg) per day at 11 weeks postpartum.⁵⁷ However, caloric requirements are only partially met by feed consumption in the first weeks postpartum. Consequently, high-producing dairy cows undergo NEB during the first four to six weeks postpartum, which often averages -5 Mcal NE_L/day, the equivalent of approximately 2.2 lb (1 kg) of body weight loss per day, mostly from adipose tissue. Reduced circulating concentrations of glucose and insulin up-regulate the lipolytic signals that result in hydrolysis of stored triglycerides from the adipose tissue, and increase the availability of non-esterified fatty acids (NEFA) to be used as an energy source. Some of the NEFA are removed by the liver, and uptake of NEFA depends on the type of fatty acid present in the circulation.⁴⁸ When the uptake of NEFA by the hepatic tissue is excessive, then re-esterification to triglycerides and ketogenesis in the hepatocytes increase.

Energy balance in early lactation has been positively associated with reproductive performance of dairy cows.⁷ The severity and length of NEB can be estimated through changes in body condition score (BCS). Cows

Table 1. Association among clinical and subclinical diseases and fertility responses in dairy cows.

Health problem	Estrous cyclic (%) [*]	AOR (CI) ¹	P
Healthy	95.6 ^a	1.00	---
Subclinical disease only	88.9 ^{b,c}	0.35 (0.16-0.76)	<0.01
Clinical disease only	93.0 ^{a,b}	0.63 (0.23-1.75)	0.37
Subclinical and clinical disease	83.5 ^c	0.23 (0.10-0.50)	<0.01
Health problem	Pregnant d 30 (%) ^{*,¶}	AOR (CI)	P
Healthy	73.5 ^a	1.00	---
Subclinical disease only	63.1 ^b	0.67 (0.44-0.99)	0.05
Clinical disease only	54.8 ^{b,c}	0.44 (0.26-0.75)	<0.01
Subclinical and clinical disease	50.0 ^c	0.39 (0.24-0.61)	<0.01
Health problem	Pregnant d 65 (%) ^{*,¶}	AOR (CI)	P
Healthy	66.2 ^a	1.00	---
Subclinical disease only	57.1 ^{a,b}	0.72 (0.49-1.05)	0.09
Clinical disease only	46.3 ^{b,c}	0.45 (0.26-0.76)	<0.01
Subclinical and clinical disease	42.1 ^c	0.39 (0.25-0.61)	<0.01

Data from Ribeiro *et al*⁵⁸

¹AOR = adjusted odds ratio; CI = confidence interval.

^{a,b,c}Superscripts within a day of pregnancy differ ($P < 0.07$).

Contrasts: * Effect of uterine disease (Healthy vs. all others) $P < 0.05$; ¶ Additive effect of metritis and clinical endometritis (clinical endometritis only + metritis only vs. metritis and clinical endometritis) $P < 0.05$.

that lost more body condition during the first 65 days postpartum were more likely to be anovular at the end of the voluntary waiting period, had decreased pregnancy per AI, and increased risk of pregnancy loss after the first AI postpartum.⁶³ Using circulating concentration of NEFA as an indicator of the energetic status of grazing dairy cows in the first two weeks postpartum, Ribeiro *et al*⁵⁸ showed that cows under NEB (NEFA ≥ 0.7 mM) were less likely to resume ovarian cyclicity before 50 days postpartum and to become pregnant to the first AI of the breeding season. Others have reported similar results in dairy herds managed under confinement. The rate of pregnancy in the first 70 days of breeding was 16% less for cows with blood NEFA ≥ 0.7 mM than for those with concentrations below this threshold in early lactation.⁵¹ Ketosis resultant from extensive fat mobilization has also been associated with compromised fertility. Both the relative circulating concentration of β -OH-butyrate (BHBA) and the duration of elevated BHBA concentrations were negatively associated with the probability of pregnancy following the first postpartum AI.⁷⁴ In fact, for every 100 μ M increase in BHBA concentration in weeks 1 and 2 after calving, the proportion of pregnant cows at first AI was reduced by 2 and 3%, respectively. Furthermore, the rate of pregnancy within 70 days after the end of the voluntary waiting period was 13% lower among cows with blood BHBA $\geq \sim 962$ μ M compared with herd mates with concentrations below 962 μ M.⁵¹ There-

fore, circulating concentrations of these metabolites can be used as indicators of excessive lipid mobilization that interfere with fertility. Furthermore, as the prevalence of cows with elevated concentrations of blood NEFA or BHBA increases, reproductive performance declines.⁵² In the latter study, the 21-day cycle pregnancy rate was reduced by 0.9 percentage units in herds in which more than 15% of the sampled cows had NEFA ≥ 0.7 mM, and by 0.8 percentage units if more than 15% of the sampled cows had BHBA ≥ 1150 μ M.

The reduction in fertility associated with low nutrient intake and NEB is, at least in part, mediated by the damaging effects on immunity and postpartum health. Exposing immune cells *in vitro* to NEFA at concentrations compatible with those observed in high-producing postpartum dairy cows (0.12 to 1 mM) has been shown to reduce function and viability. Increasing the concentration of NEFA in the culture media abridged the synthesis of interferon- γ and IgM by peripheral blood mononuclear cells.³⁸ Furthermore, NEFA reduced phagocytosis-dependent oxidative burst in polymorphonuclear leukocytes.⁶⁹ When concentrations of NEFA in the culture medium were further increased to 2 mM, polymorphonuclear oxidative burst was not altered, but more leukocytes underwent necrosis, thereby impairing function. Not only NEFA, but also BHBA has been implicated with immunosuppression in postpartum dairy cows. Incubation of bovine neutrophils with increasing

concentrations of BHBA reduced phagocytosis, extracellular trap formation, and killing of bacteria.²⁸ *In vivo* observations support the immunosuppressive effects of NEB. Cows under severe NEB had increased NEFA and BHBA, which was associated with decreased leukocyte numbers.⁷⁵ It is likely that cows unable to recover feed consumption after parturition and, therefore remain in more severe NEB, are more susceptible to diseases. It is known that reduced nutrient intake and NEB even before calving are associated with poor uterine recovery from parturition and the occurrence of uterine diseases.³¹ These observations seem to be linked with changes in patterns of endometrium gene expression mediated by the energetic status of the cows. Wathes *et al*⁷⁵ evaluated global gene expression of the endometrium of cows at two weeks postpartum. They observed that several probes linked with inflammation and active immune response were still up-regulated in cows undergoing severe NEB compared with those exhibiting a more modest caloric deficit, suggesting a delay in uterine involution. In addition, cows that developed uterine diseases in early postpartum had greater concentrations of NEFA and BHBA around calving than healthy cows.^{22,31} It is important to highlight that the occurrence of diseases early postpartum can further accentuate the adverse effects of NEB, as sick cows have reduced appetite and oftentimes lose more body weight than healthy cows.

In addition to the changes in energy balance, circulating concentrations of antioxidants such as β -carotene, and vitamins A (retinol) and E (α -tocopherol) are also temporally regulated and decrease around parturition.²⁵ As these compounds play important roles in immune function, low concentrations of these vitamins have been associated with increased susceptibility to disease and, potentially, with reduced fertility in dairy cows. Prepartum circulating β -carotene and, more importantly, vitamin E were lower for cows that retained their placenta than for healthy cows.³⁹ In fact, for every 1 μ g/mL increase in circulating vitamin E during the week preceding parturition, the risk of retained placenta decreased by 21%. Furthermore, the decline in circulating concentrations of β -carotene, vitamin A, and vitamin E associated with parturition was more accentuated among cows that developed mastitis during the first 30 days postpartum than among healthy cows.³⁹ In the last week prepartum, a 100 ng/mL increase in circulating vitamin A concentration was associated with a 60% decrease in the risk of clinical mastitis.³⁹

Impact of Energy Balance on Oocyte Competence and Early Embryo Development

During lactation, most glucose produced by the liver is used for synthesis of lactose to support milk production. A transient insulin resistance early postpartum dimin-

ishes the utilization of glucose by peripheral tissues to secure its availability to the mammary gland. Although the follicle is capable of controlling fluctuations in glucose availability, which generally results in concentrations in the follicular fluid greater than those observed in blood, intra-follicular glucose concentrations also decline around parturition.⁴² It has been shown that glucose is critical for adequate oocyte maturation, affecting cumulus expansion, nuclear maturation, cleavage, and subsequent blastocyst development. In fact, glucose concentrations compatible with those observed in cows suffering from clinical ketosis (1.4 mM) were shown to reduce cleavage and the proportion of embryos developing to blastocysts.⁴⁰ Although the oocyte does not directly use glucose as an energy source, it has to be readily available for cumulus cells for glycolysis to provide pyruvate and lactate, oocyte's preferred substrates for ATP production.¹¹ Therefore, it is possible that hypoglycemia in early lactation might compromise oocyte competence in dairy cows.

Extensive fat mobilization and the release of large amounts of NEFA into the bloodstream have been shown to exert a direct effect on fertility of postpartum dairy cows. Concentrations of NEFA in the follicular fluid parallel those of serum, and they increase around parturition.⁴¹ Maturation of oocytes *in vitro* in the presence of saturated fatty acids reduced oocyte competence and compromised the initial development of embryos. Specifically, the addition of palmitic and stearic acids to the maturation media induced apoptosis and necrosis of cumulus cells, which was associated with impaired fertilization, cleavage, and development to the blastocyst stage.⁴¹ Changes in circulating concentrations of BHBA are promptly reflected in the follicular fluid.⁴² However, *in vitro* models developed to study the effects of subclinical ketosis on fertility of dairy cows have failed to demonstrate a direct effect of BHBA on oocyte competence, which seems only to aggravate the responses to low concentrations of glucose during oocyte maturation.⁴⁰

Energy Balance and Ovarian Function Postpartum

The stage set by NEB modulates the activity of the hypothalamic-pituitary-ovarian axis. Under-nutrition has been linked to the inability of the hypothalamus to sustain high frequency of LH pulses by the pituitary gland.⁷⁰ Indeed, LH pulse frequency was shown to be positively correlated with energy balance and negatively correlated with blood NEFA.³⁴ The underlying mechanism by which NEB reduces LH release is likely to involve supply of oxidizable fuels to neurons and hormonal modulation of hypothalamic and pituitary cells.⁷¹ Glucose is a preferred substrate for neuron energy metabolism, and inadequate supply of glucose inhibits the GnRH pulse generator.⁷¹ Under a favorable nutritional status, the hormonal mi-

lieu to which the hypothalamus and pituitary gland are exposed favors the release of GnRH and gonadotropins. For instance, leptin, a hormone known to have increased concentrations under positive energy balance, stimulates the release of GnRH by the hypothalamus, and blood leptin was found to be strongly correlated with both LH pulse frequency and amplitude.³⁴ In addition to low LH support, cows under NEB have limited hepatic expression of GH receptor 1A triggered by low circulating concentrations of insulin.^{6,8} This phenomenon uncouples the growth hormone (GH)/insulin-like growth factor-1 (IGF-1) axis, which reduces the synthesis of IGF-1 by the liver. Reduced concentrations of IGF-1 have been associated with diminished follicle sensitivity to LH, growth and steroidogenesis.^{6,46} Conversely, the increase in circulating concentrations of insulin as energy balance improves seems to be one of the signals to re-establish the GH receptor expression in the liver and restore IGF-1 synthesis in dairy cows.⁸ Restricting follicular growth and synthesis of estradiol delay resumption of ovulation postpartum and might compromise oocyte quality, which likely hampers estrous detection and pregnancy in dairy cows.

In addition to extensive nutrient shortage, high-producing dairy cows also undergo extensive ovarian steroid catabolism. This is thought to be mediated by the high dry matter intake and consequent increased splanchnic blood flow.⁶⁰ Hepatic blood flow doubles in the first three months postpartum, averaging 1,147 liters per hour in the week preceding parturition and 2,437 liters per hour in the third month postpartum.⁵⁷ The increased clearance of ovarian steroids can have important implications to the reproductive biology of dairy cows and indirectly influence follicle development,⁷⁸ which can have implications for oocyte quality and subsequent embryo development. Progesterone-induced uterine histotroph secretion is critical for the nourishment and elongation of the bovine conceptus.⁵⁹ Therefore, an increase in the rate of progesterone clearance is expected to result in a slower rise in progesterone concentrations after insemination, reducing embryo development,⁵⁹ which has implications for pregnancy. Similarly, reduced circulating concentrations of estradiol because of hepatic catabolism in cows with high dry matter intake can result in a shorter and less intense estrus period.⁴⁵ In addition, estradiol catabolism requires follicles to grow for longer periods of time to be able to trigger estrus and ovulation.^{68,78} Longer periods of follicular dominance reduce embryo quality⁹ and pregnancy per AI in cows inseminated on estrus⁴ or following timed AI.⁶²

Calcium Homeostasis and Uterine Health during Early Postpartum

The control of blood concentrations of Ca is critical to maintain normal muscle contractility, transmission

of nerve impulses, and immune function. Nonetheless, homeostatic controls in early lactation in high-producing dairy cows might not prevent declines in Ca concentrations in the first week postpartum. The amount of Ca secreted in colostrum on the day of calving is almost 8 to 10 times the entire serum Ca pool in a dairy cow.²⁴ Therefore, it is no surprise that most cows undergo a period of subclinical hypocalcemia and a portion of them develop milk fever. In fact, surveys in the US indicate that 25, 41, 49, 51, 54, and 42% of first to sixth-lactation cows are hypocalcemic (Ca < 8 mg/dL) in the first 48 hours after calving.⁵⁶ In order to maintain serum total and ionized Ca (Ca²⁺), postpartum dairy cows have to increase bone remodeling for Ca resorption or increase intestinal Ca absorption.

The impact of milk fever on the health of dairy cows is very conspicuous, as it results in downer cows and death if left untreated. Nevertheless, milder depressions of serum Ca concentrations are often not diagnosed and have a pronounced negative effect on postpartum health and fertility. Recently, Martinez *et al*⁴⁷ observed that cows with serum Ca ≤ 8.59 mg/dL in at least one of the first three days postpartum had reduced neutrophil phagocytic and killing activities *in vitro*, increased odds of developing fever (adjusted OR = 3.5; 95% CI = 1.1-11.6) and metritis (adjusted OR = 4.5; 95% CI = 1.3-14.9), and these associations were observed for both cows considered to be of high or low risk of developing metritis based on calving problems. The authors concluded that the attributable risk for a cow to develop metritis because of low serum Ca was 75.3%.⁴⁷ Ionized Ca is an important second messenger in cellular signal transduction, fluctuations in intracellular Ca²⁺ concentrations are critical to activate immune cells,⁴³ and cows with retained placenta have reduced neutrophil function.³⁷ Intracellular stores and flux of Ca²⁺ in response to cell activation are reduced in lymphocytes of dairy cows with milk fever.³⁶ Collectively, these data suggest that Ca status is linked with immune cell function and plays a role in the risk of uterine diseases of dairy cows. Cows suffering from uterine diseases have delayed postpartum ovulation, reduced pregnancy per AI, and increased pregnancy loss.⁶¹

Management of Transition Cows to Improve Periparturient Health and Fertility

The multifactorial nature of reproduction requires a “holistic” and integrated approach to management from housing to feeding and breeding.

Cow Movement and Dry Period Length

Regrouping of cows induces social behaviors that oftentimes disturb feeding and resting patterns, thereby resulting in a temporary increase in aggression concur-

rently with a reduction in dry matter intake.⁷³ Therefore, regrouping cows at the imminence of calving is not advised, as it would further suppress intake and increase the risk of ketosis and fatty liver. However, the question of when cows can and cannot be moved still remains. Recent work from Wisconsin refuted the concept that weekly addition of cows to the close-up group is detrimental to postpartum metabolism and production.¹³ It seems that when appropriate feedbunk space and number of stalls are available, transition cows can adapt to the weekly regrouping.

A strategy to improve postpartum intermediary metabolism is to manipulate the length of the dry period. Reducing the dry period from 55 to 34 days increased BCS between weeks 2 and 8 postpartum and reduced the concentrations of plasma NEFA at week 3 postpartum,⁷⁷ suggesting improved energy status in early lactation. When energy balance was measured, cows subjected to a 28-day dry period experienced a less severe NEB postpartum, which resulted in reduced BCS and body weight losses compared with cows having the traditional 56 days dry.⁵⁵ Some of the benefit to energy balance is the result of less milk production, particularly in cows starting their second lactation.^{66,77} The improved energy balance with a short dry period likely explains the earlier first postpartum ovulation and reduction in anovular cows.^{29,76} Despite changes in energy status and an earlier resumption of estrous cyclicity, cows with a dry period of 28 to 35 days had similar reproductive performance to those with a standard eight-week dry period.^{29,67,76} Nevertheless, in observational studies, extending the exposure of cows to the prepartum diet was associated with reduced days open and increased proportion of pregnant cows at weeks 6 and 21 after the initiation of the breeding season.¹⁵

Prepartum Diet Formulation

Altering caloric intake prepartum influences postpartum metabolism in dairy cows. *Ad libitum* nutrient intake during the entire dry period tends to increase body weight and BCS prepartum and predispose cows to increased lipid mobilization in early lactation.¹⁸ Several studies have evaluated the impact of manipulating the energy density of the prepartum diet on postpartum performance. In some cases, nutrient intake was restricted not by altering the diet formulation, but by limiting the amount of feed offered. Tables 2 and 3 depict a summary of studies in the literature in which caloric intake was restricted during the last weeks of gestation and the impacts on subsequent lactation fat-corrected milk and blood ketones. In general, restricting nutrient intake resulted in an average reduction of 4.4 lb (2 kg) per day of fat-corrected milk, with minor effects on BHBA concentrations. In some studies, high caloric intake resulted in greater triacylglycerol accumulation in the liver^{18,32} because of greater fat mobilization measured as plasma NEFA. The increased postpartum lipid mobilization is likely the result of increased milk yield without a concurrent increase in dry matter intake. Therefore, restricting caloric intake prepartum can be used to minimize lipid mobilization and triacylglycerol accumulation in the liver, but at the expense of milk production.

Altering the protein content of the prepartum diet has little impact on performance of postpartum multiparous cows; however, increasing prepartum dietary protein from 12.7 to 14.7% of the diet dry matter with a high rumen undegradable protein source enhanced milk production in primiparous cows.⁶⁵ Nonetheless, protein had negligible impacts on measures of reproduction. Time to resumption of ovulation postpartum, days open, and pregnancy per AI were all unaffected by

Table 2. Effect of prepartum caloric intake on fat-corrected milk (kg/d).

Reference	Prepartum intake ¹		P ²
	Low caloric intake	High caloric intake	
Douglas <i>et al</i> ¹⁹	35.6	37.9	NS
Douglas <i>et al</i> ¹⁸	40.8	39.8	NS
Rabelo <i>et al</i> ^{53,54}	38.5	40.4	0.59
Doepel <i>et al</i> ¹⁷	39.1	40.3	NS
Hayirli <i>et al</i> ³⁰	33.7	35.2	0.27
Janovick and Drackley ³²	40.5	46.1	0.09
Kanjanapruthipong <i>et al</i> ³⁵	26.1	28.4	0.04
Average	36.3	38.3	

¹Prepartum caloric intake (net energy for lactation) averaged 14.6 and 19.8 Mcal of net energy for lactation/cow/day for the low and the high caloric intake, respectively.

²NS = not significant ($P > 0.10$).

Table 3. Effect of prepartum caloric intake on plasma/serum concentrations of β -OH-butyrate (mg/dL).

Reference	Prepartum intake ¹		<i>P</i> ²
	Low caloric intake	High caloric intake	
Douglas <i>et al</i> ¹⁹	5.2	4.7	0.56
Douglas <i>et al</i> ¹⁸	4.8	6.0	0.05
Rabelo <i>et al</i> ^{53,54}	5.4	5.0	0.45
Doepel <i>et al</i> ¹⁷	~10	~10	NS
Hayirli <i>et al</i> ³⁰	11.6	11.4	0.96
Janovick and Drackley ³²	4.5	6.6	0.01
Kanjanapruthipong <i>et al</i> ³⁵	6.1	3.8	0.04
Average	6.8	6.8	

¹Prepartum caloric intake (net energy for lactation) averaged 14.6 and 19.8 Mcal of net energy for lactation/cow/day for the low and the high caloric intake, respectively.

²NS = not significant ($P > 0.10$).

prepartum dietary protein concentration. Similarly, the incidence of diseases postpartum was not affected by prepartum dietary protein. Therefore, diets for cows in the last weeks of gestation should contain between 12% (multiparous cows) and 15% (primigravid cows) crude protein to result in an estimated 0.45 lb (1 kg) per day of metabolizable protein intake.⁴⁹

Increasing Postpartum Blood Insulin

A number of studies have demonstrated the importance of insulin as a signal mediating the effects of acute changes in nutrient intake on reproductive parameters in dairy cattle. Feeding more dietary starch or enhancing the ruminal fermentability of starch in the diet usually results in increased plasma insulin concentrations. Insulin mediates recoupling of the GH/IGF-1 axis,⁸ which is important for follicle development and ovulation. Gong *et al*²⁷ fed cows of low- and high-genetic merit isocaloric diets that differed in the ability to induce high or low insulin concentrations in plasma. Feeding the high-starch diet reduced the interval to first postpartum ovulation and resulted in a greater proportion of estrous cyclic cows within the first 50 days postpartum. Nevertheless, this response has not been consistent.²³ It is important to remember that although diets high in starch favor increases in plasma insulin, excessive amounts of readily fermentable starch has the potential to suppress dry matter intake and offset any potential benefits of dietary manipulation on ovarian function.

Altering Hepatic Lipid Metabolism

During periods of extensive fat mobilization, fat accumulates in the hepatic tissue. In early lactation cows with relatively low plasma NEFA concentrations (0.36 mM), the liver extracted 724 g of NEFA from blood over

a 24-hour period.⁵⁷ Thus, in cows with concentrations of NEFA above 1 mM, as those with extensive lipid mobilization immediately after calving, the liver might remove as much as 4.4 lb (2 kg) of NEFA per day, the equivalent of 20% of its weight. Most of these NEFA reaching the liver are oxidized for energy production or converted into BHBA, with a smaller contribution for synthesis of very low-density lipoprotein (VLDL). The bovine liver has limited capacity to synthesize and secrete VLDL, thereby compromising export of triacylglycerols during periods of extensive hepatic NEFA uptake. The resulting hepatic lipidosis has been associated with retained placenta, ketosis, displaced abomasum, and impaired immune function and reproduction.^{5,33} Thus, reducing the risk of lipid-related disorders might improve reproduction of dairy cows. Supplementation of periparturient dairy cows with rumen-protected choline has been used as a strategy to improve lipid metabolism and alleviate hepatic lipidosis. When feed intake was restricted to 30% of the maintenance to simulate a period of NEB and induce hepatic lipidosis, the supplementation of rumen-protected choline reduced triacylglycerol accumulation in the liver.¹² Furthermore, the inclusion of supplemental choline in the diet from approximately 25 days before to 80 days after calving reduced loss of body condition postpartum and concentrations of BHBA, which resulted in lower incidence of clinical and subclinical ketosis despite the increase in fat-corrected milk.⁴⁴ Although feeding rumen-protected choline reduced morbidity and improved metabolic health, no benefits were observed for reproduction. Supplemental rumen-protected choline did not affect the resumption of postpartum estrous cyclicity, pregnancy per AI at the first and second inseminations, or maintenance of pregnancy in the first 60 days of gestation.

Supplementing Ionophores to Periparturient Dairy Cows

Ionophores are lipophilic molecules involved with ionic transport across cell membranes. Monensin is a carboxylic polyether ionophore that has been used in animal nutrition because it selectively inhibits gram-positive bacteria. The shift in the rumen microbiota caused by monensin favors propionate production and N conservation by reducing ruminal proteolysis. Feeding monensin typically increases blood glucose and insulin and reduces NEFA and BHBA.²⁰ In association with improved metabolic health, monensin was effective in reducing the incidence of ketosis, displaced abomasum, and mastitis.²¹ When monensin was supplemented as a controlled-release capsule, it reduced the incidence of metritis.²¹ Surprisingly, feeding monensin to dairy cows during the transition period has not been shown to hasten resumption of ovulation postpartum, to reduce days to pregnancy, or to increase the rate of pregnancy, in spite of consistent improvements in metabolic health.^{1,21}

Improving Ca Homeostasis Postpartum

Improving serum concentrations of Ca in early lactation is achieved by enhancing bone mineral resorption, intestinal absorption of dietary Ca, and by increasing the ionized Ca fraction in blood. A common method to improve Ca homeostasis is to manipulate the dietary cation-anion difference (DCAD) prepartum.^{24,26,72} Reducing the DCAD by feeding salts with strong anions decreases blood pH and enhances the affinity of the parathyroid hormone (PTH) to the PTH receptor present on cells in the bones, intestine, and kidneys.²⁴ Although feeding strong anions reduces feed intake during supplementation, the improved postpartum Ca metabolism often results in greater postpartum feed intake.¹⁶ Feeding acidogenic diets prepartum did not reduce the incidences of retained placenta, lameness, and subclinical ketosis.⁷² However, supplementing cows with calcium chloride in a gel formulation 12 hours before the expected calving and at 0, 12, and 24 hours after calving reduced the incidence of clinical and subclinical hypocalcemia, and displacement of abomasum.⁵⁰ Despite the benefits of feeding acidogenic diets on Ca homeostasis and the link between serum Ca and uterine diseases and reproduction in dairy cows,⁴⁷ intervals to first insemination and pregnancy were not affected by feeding a low DCAD diet prepartum.⁷² Additional research is needed with properly powered experiments to critically evaluate the impact of reducing subclinical hypocalcemia by manipulating the DCAD of prepartum diets or supplementing Ca postpartum on reproduction of dairy cows.

Fatty Acid Supplementation and Reproduction of Dairy Cows

Fatty acids (FA) are structural and functional lipid components of cells capable of modulating membrane

fluidity and permeability, and carrying out cell signaling, in addition to providing calories for tissue energy metabolism. Therefore, fat supplementation to dairy cattle diets provides benefits that go beyond the provision of calories, and these effects are influenced by the profile of FA of the fat source.

The prostaglandin (PG) $F_{2\alpha}$ synthesized by the endometrium plays an important role in reproduction of dairy cows. Dietary FA supplementation has been shown to influence tissue FA composition, which makes it possible to manipulate arachidonic acid concentration in the endometrium, the precursor for $PGF_{2\alpha}$ synthesis.² In fact, the feeding of supplemental fat prepartum (30% of FA as C18:2 n-6) enhanced uterine secretion of $PGF_{2\alpha}$.¹⁴ Prepartum supplementation with Ca salts of long chain FA rich in n-6 FA reduced the incidence of retained placenta, metritis, and mastitis compared with cows not fed fat prepartum.¹⁴ Similarly, supplementing prepartum diets with 2% Ca salts of either palm oil or a blend of C18:2 n-6 and trans-octadecenoic FA reduced the severity of uterine disease postpartum.⁶⁴

Although the incorporation of supplemental fat enriched in polyunsaturated FA has been shown to influence follicle growth,⁶⁴ it is unclear whether supplemental fats differing in FA profile have any effect on resumption of cyclicity. In general, type of FA does not change the proportion of estrous cyclic cows at around 60 days postpartum.⁶⁴ Even though the amount of lipids in the oocyte of ruminants is greater than that in most studied species, which would lead to the potential to change FA profile to influence oocyte competence, feeding polyunsaturated FA did not impact oocyte quality based on subsequent embryo development *in vitro*.³ Nevertheless, studies *in vivo* support the concept that altering the FA profile of the diet influences fertilization and embryo quality in lactating dairy cows.^{10,64} Collectively, results from studies on the effects of fat supplementation on fertility of dairy cows suggest that incorporating fat into dairy cattle rations improves pregnancy per AI.⁶⁴ Furthermore, the type of FA seems to play a role in the establishment and maintenance of pregnancy in cattle.⁶⁴

Conclusions

It is accepted that reproduction is important for the profitability of dairy farms, and metabolic health is associated with successful reproduction. Cows that experience periparturient problems have delayed return to ovulation, lower pregnancy per insemination, and increased pregnancy loss. Therefore, implementing nutritional and health programs that reduce the risk of metabolic disturbances are expected to not only improve cow health, but also enhance fertility. Strategies to manipulate peripartum metabolic health involve dietary formulation to minimize the degree and extent

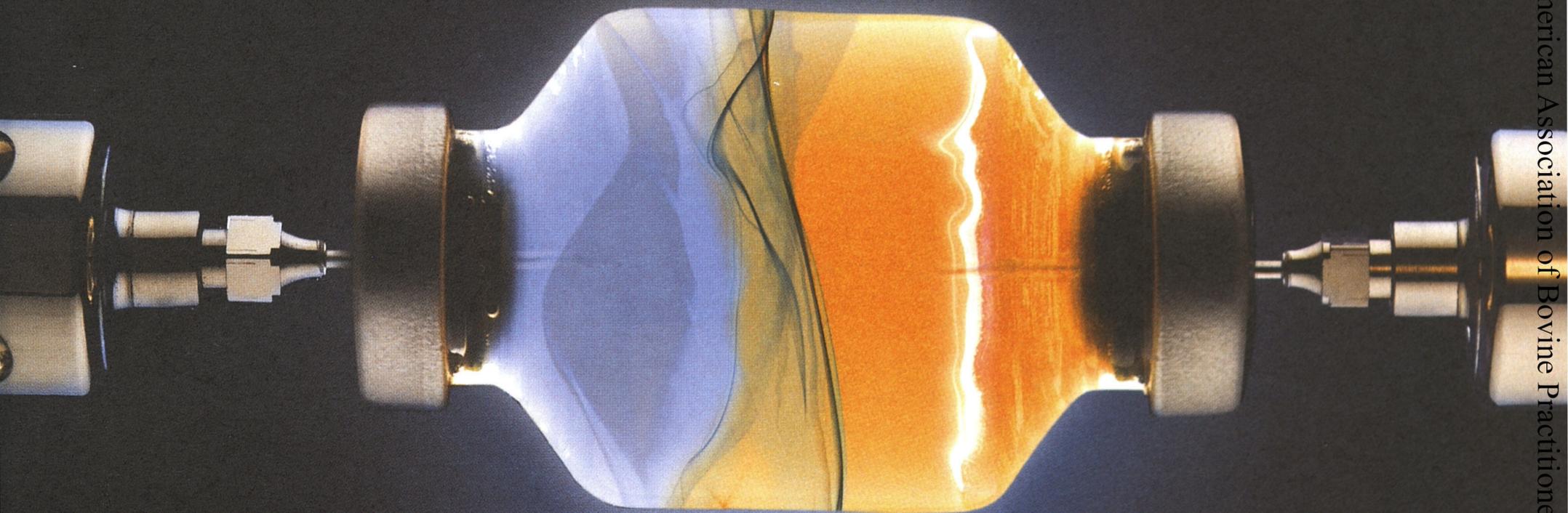
of negative nutrient balance, improve Ca homeostasis, and minimize the severity of immunosuppression around and immediately after calving.

References

1. Abe N, Lean IJ, Rabiee A, Porter J, Graham C: Effects of sodium monensin on reproductive performance of dairy cattle. II. Effects on metabolites in plasma, resumption of ovarian cyclicity and oestrus in lactating cows. *Aust Vet J* 71:277-282, 1994.
2. Bilby TR, Guzeloglu A, MacLaren LA, Staples CR, Thatcher WW: Pregnancy, bST and omega-3 fatty acids in lactating dairy cows: II. Gene expression related to maintenance of pregnancy. *J Dairy Sci* 89:3375-3385, 2006.
3. Bilby TR, Block J, do Amaral BC, Sa Filho O, Silvestre FT, Hansen PJ, Staples CR, Thatcher WW: Effects of dietary unsaturated fatty acids on oocyte quality and follicular development in lactating dairy cows in summer. *J Dairy Sci* 89:3891-3903, 2006.
4. Bleach ECL, Glencross RG, Knight PG: Association between ovarian follicle development and pregnancy rates in dairy cows undergoing spontaneous oestrous cycle. *Repro* 127:621-629, 2004.
5. Bobe G, Young JW, Beitz DC: Invited review: pathology, etiology, prevention, and treatment of fatty liver in dairy cows. *J Dairy Sci* 87, 3105-3124, 2004.
6. Butler ST, Pelton SH, Butler WR: Insulin increases 17 beta-estradiol production by the dominant follicle of the first postpartum follicle wave in dairy cows. *Repro* 127:537-545, 2004.
7. Butler WR: Energy balance relationships with follicular development, ovulation and fertility in postpartum dairy cows. *Livest Prod Sci* 83:211-218, 2003.
8. Butler ST, Marr AL, Pelton SH, Radcliff RP, Lucy MC, Butler WR: Insulin restores GH responsiveness during lactation-induced negative energy balance in dairy cattle: effects on expression of IGF-I and GH receptor 1A. *J Endocrinol* 176:205-217, 2003.
9. Cerri RL, Rutigliano HM, Chebel RC, Santos JEP: Period of dominance of the ovulatory follicle influences embryo quality in lactating dairy cows. *Repro* 137:813-823, 2009.
10. Cerri RLA, Juchem SO, Chebel RC, Rutigliano HM, Bruno RGS, Galvão KN, Thatcher WW, Santos JEP: Effect of fat source differing in fatty acid profile on metabolic parameters, fertilization, and embryo quality in high-producing dairy cows. *J Dairy Sci* 92:1520-1531, 2009.
11. Cetica P, Pintos L, Dalvit G, Beconi M: Activity of key enzymes involved in glucose and triglyceride catabolism during bovine oocyte maturation in vitro. *Repro* 124:675-681, 2002.
12. Cooke RF, Silva Del Río N, Caraviello DZ, Bertics SJ, Ramos MH, Grummer RR: Supplemental choline for prevention and alleviation of fatty liver in dairy cattle. *J Dairy Sci* 90:2413-2418, 2007.
13. Coonen JM, Maroney MJ, Crump PM, Grummer RR: Short communication: Effect of a stable pen management strategy for precalving cows on dry matter intake, plasma nonesterified fatty acid levels, and milk production. *J Dairy Sci* 94:2413-2417, 2001.
14. Cullens FM, Staples CR, Bilby TR, Silvestre FT, Bartolome J, Sozzi A, Badinga L, Thatcher WW, Arthington JD: Effect of timing of initiation of fat supplementation on milk production, plasma hormones and metabolites, and conception rates of Holstein cows in summer. *J Dairy Sci* 86 (Suppl. 1):308 (Abstract), 2004.
15. DeGaris PJ, Lean IJ, Rabiee AR, Heuer C: Effects of increasing days of exposure to prepartum transition diets on reproduction and health in dairy cows. *Aust Vet J* 88:84-92, 2010.
16. DeGroot MA, Block E, French PD: Effect of prepartum anionic supplementation on periparturient feed intake, health, and milk production. *J Dairy Sci* 93:5268-5279, 2010.
17. Doepel L, Lapierre H, Kennelly JJ: Peripartum performance and metabolism of dairy cows in response to prepartum energy and protein intake. *J Dairy Sci* 85:2315-2334, 2002.
18. Douglas GN, Overton TR, Bateman II HG, Dann HM, Drackley JK: Prepartal plane of nutrition, regardless of dietary energy source, affects periparturient metabolism and dry matter intake in Holstein cows. *J Dairy Sci* 89:2141-2157, 2006.
19. Douglas GN, Rehage J, Beaulieu AD, Bahaa AO, Drackley JK: Prepartum nutrition alters fatty acid composition in plasma, adipose tissue, and liver lipids of periparturient dairy cows. *J Dairy Sci* 90:2941-2959, 2007.
20. Duffield TF, Rabiee AR, Lean IJ: A meta-analysis of the impact of monensin in lactating dairy cattle. Part 1. Metabolic effects. *J Dairy Sci* 91:1334-1346, 2008.
21. Duffield TF, Rabiee AR, Lean IJ: A meta-analysis of the impact of monensin in lactating dairy cattle. Part 2. Health and reproduction. *J Dairy Sci* 91:2328-2341, 2008.
22. Galvão KN, Flaminio MJB, Brittin SB, Sper R, Fraga M, Caixeta L, Ricci A, Guard CL, Butler WR, Gilbert RO: Association between uterine disease and indicators of neutrophil and systemic energy status in lactating Holstein cows. *J Dairy Sci* 93:2926-2937, 2010.
23. Garnsworthy PC, Fouladi-Nashta AA, Mann GE, Sinclair KD, Webb R: Effect of dietary-induced changes in plasma insulin concentrations during the early post partum period on pregnancy rate in dairy cows. *Repro* 137:759-768, 2009.
24. Goff JP: Macromineral disorders of the transition cow. *Vet Clin North Am Food Anim Pract* 20:471-494, 2004.
25. Goff JP, Kimura K, Horst RL: Effect of mastectomy on milk fever, energy, and vitamins A, E, and β -carotene at parturition. *J Dairy Sci* 85:1427-1436, 2002.
26. Goff JP, Horst RL, Mueller FJ, Miller JK, Kiess GA, Dowlen HH: Addition of chloride to a prepartal diet high in cations increases 1,25-dihydroxyvitamin D response to hypocalcemia preventing milk fever. *J Dairy Sci* 74:3863-3871, 1991.
27. Gong JG, Lee WJ, Garnsworthy PC, Webb R: Effect of dietary-induced increases in circulating insulin concentrations during the early postpartum period on reproductive function in dairy cows. *Reprod* 123:419-427, 2002.
28. Grinberg N, Elazar S, Rosenshine I, Shpigel NY: β -hydroxybutyrate abrogates formation of bovine neutrophil extracellular traps and bactericidal activity against mammary pathogenic *Escherichia coli*. *Infect Immun* 76:2802-2807, 2008.
29. Gümen A, Rastani RR, Grummer RR, Wiltbank MC: Reduced dry periods and varying prepartum diets alter postpartum ovulation and reproductive measures. *J Dairy Sci* 88:2401-2411, 2005.
30. Hayirli A, Keisler DH, Doepel L, Petit H: Peripartum responses of dairy cows to prepartal feeding level and dietary fatty acid source. *J Dairy Sci* 94:917-930, 2011.
31. Hammon DS, Evjen IM, Dhiman TR, Goff JP, Walters JL: Neutrophil function and energy status in Holstein cows with uterine health disorders. *Vet Immunol Immunopathol* 113:21-29, 2006.
32. Janovick NA, Drackley JK: Prepartum dietary management of energy intake affects postpartum intake and lactation performance by primiparous and multiparous Holstein cows. *J Dairy Sci* 93:3086-3102, 2010.
33. Jorritsma R, Jorritsma H, Schukken YH, Wentink GH: Relationships between fatty liver and fertility and some periparturient diseases in commercial Dutch dairy herds. *Therio* 54:1065-1074, 2000.
34. Kadokawa H, Blache D, Martin GB: Plasma leptin concentrations correlate with luteinizing hormone secretion in early postpartum Holstein cows. *J Dairy Sci* 89:3020-3027, 2006.
35. Kanjanapruthipong J, Homwong N, Buatong N: Effects of prepartum roughage neutral detergent fiber levels on periparturient dry matter intake, metabolism, and lactation in heat-stressed dairy cows. *J Dairy Sci* 93:2589-2597, 2010.
36. Kimura K, Reinhardt TA, Goff JP: Parturition and hypocalcemia blunts calcium signals in immune cells of dairy cattle. *J Dairy Sci* 89:2588-2595, 2006.
37. Kimura K, Goff JP, Kehrli ME Jr, Reinhardt TA: Decreased neutrophil function as a cause of retained placenta in dairy cattle. *J Dairy Sci* 85:544-550, 2002.
38. Lacetera N, Scalia D, Franci O, Bernabucci U, Ronchi B, Nardone A: Short communication: effects of non-esterified fatty acids on lymphocyte function in dairy heifers. *J Dairy Sci* 87:1012-1014, 2004.
39. LeBlanc SJ, Herdt TH, Seymour WM, Duffield TF, Leslie KE: Peripartum serum vitamin E, retinol, and beta-carotene in dairy cattle and their associations with disease. *J Dairy Sci* 87:609-619, 2004.

40. Leroy JLMR, Vanholder T, Opsomer G, Van Soom A, de Kruif A: The in vitro development of bovine oocytes after maturation in glucose and b-hydroxybutyrate concentrations associated with negative energy balance in dairy cows. *Reprod Dom Anim* 41:119-123, 2006.
41. Leroy JLMR, Vanholder T, Mateusen B, Christophe A, Opsomer G, de Kruif A, Genicot G, Van Soom A: Non-esterified fatty acids in follicular fluid of dairy cows and their effect on developmental capacity of bovine oocytes in vitro. *Repro* 130:485-495, 2005.
42. Leroy JLMR, Vanholder T, Delanghe JR, Opsomer G, Van Soom A, Bols PEJ, Dewulf J, de Kruif A: Metabolic changes in follicular fluid of the dominant follicle in high-yielding dairy cows early post partum. *Therio* 62:1131-1143, 2004.
43. Lewis RS: Calcium signaling mechanisms in T lymphocytes. *Annu Rev Immunol* 19:497-521, 2001.
44. Lima FS, Sá Filho MF, Greco LF, Santos JEP: Effects of feeding rumen-protected choline on incidence of diseases and reproduction of dairy cows. *Vet J* (accepted in press), 2011.
45. Lopez H, Satter LD, Wiltbank MC: Relationship between level of milk production and estrous behavior of lactating dairy cows. *Anim Reprod Sci* 81:209-223, 2004.
46. Lucy MC, Beck J, Staples CR, Head HH, De La Sota RL, Thatcher WW: Follicular dynamics, plasma metabolites, hormones and insulin-like growth factor I (IGF-I) in lactating cows with positive or negative energy balance during the preovulatory period. *Reprod Nutr Dev* 32:331-341, 1992.
47. Martinez N, Risco CA, Maunsell F, Galvão KN, Santos JEP: Evaluation of periparturient calcium status and neutrophil function of dairy cows of low or high risk of developing uterine diseases. *Proc Am Assoc Bov Pract Conf* 44:151, 2011.
48. Mashek DG, Grummer RR: Short communication: Net uptake of nonesterified long chain fatty acids by the perfused caudate lobe of the caprine liver. *J Dairy Sci* 86:1218-1220, 2003.
49. NRC: *Nutrient Requirements of Dairy Cattle*, ed 7. National Research Council, Washington, DC, 2001.
50. Oetzel GR: Effect of calcium chloride gel treatment in dairy cows on incidence of periparturient diseases. *J Am Vet Med Assoc* 209:958-961, 1996.
51. Ospina PA, Nydam DV, Stokol T, Overton TR: Associations of elevated nonesterified fatty acids and β -hydroxybutyrate concentrations with early lactation reproductive performance and milk production in transition dairy cattle in the northeastern United States. *J Dairy Sci* 93:1596-1603, 2010.
52. Ospina PA, Nydam DV, Stokol T, Overton TR: Association between the proportion of sampled transition cows with increased nonesterified fatty acids and β -hydroxybutyrate and disease incidence, pregnancy rate, and milk production at the herd level. *J Dairy Sci* 93:3595-3601, 2010.
53. Rabelo E, Rezende RL, Bertics SJ, Grummer RR: Effects of transition diets varying in dietary energy density on lactation performance and ruminal parameters of dairy cows. *J Dairy Sci* 86:916-925, 2003.
54. Rabelo E, Rezende RL, Bertics SJ, Grummer RR: Effects of pre- and postfresh transition diets varying in dietary energy density on metabolic status of periparturient dairy cows. *J Dairy Sci* 88:4375-4383, 2005.
55. Rastani RR, Grummer RR, Bertics SJ, Gümen A, Wiltbank MC, Mashek DG, Schwab MC: Reducing dry period length to simplify feeding transition cows: milk production, energy balance, and metabolic profiles. *J Dairy Sci* 88:1004-1014, 2005.
56. Reinhardt TA, Lippolis JD, McCluskey BJ, Goff JP, Horst RL: Prevalence of subclinical hypocalcemia in dairy herds. *Vet J* 188:122-124, 2011.
57. Reynolds CK, Aikman PC, Lupoli B, Humphries DJ, Beaver DE: Splanchnic metabolism of dairy cows during the transition from late gestation through early lactation. *J Dairy Sci* 86:1201-1217, 2003.
58. Ribeiro ES, Lima FS, Ayres H, Greco LF, Bisinotto RS, Favoreto M, Marsola RS, Monteiro APA, Thatcher WW, Santos JEP: Effect of postpartum diseases on reproduction of grazing dairy cows. *J Dairy Sci* 94(E-Suppl. 1), 63 (Abstr.), 2011.
59. Robinson RS, Fray MD, Wathes DC, Lamming GE, Mann GE: In vivo expression of interferon tau mRNA by the embryonic trophoblast and uterine concentrations of interferon tau protein during early pregnancy in the cow. *Mol Reprod Dev* 73:470-474, 2006.
60. Sangsritavong S, Combs DK, Sartori R, Armentano LE, Wiltbank MC: High feed intake increases liver blood flow and metabolism of progesterone and estradiol-17 β in dairy cattle. *J Dairy Sci* 85:2831-2842, 2002.
61. Santos JEP, Bisinotto RS, Ribeiro ES, Lima FS, Greco LF, Staples CR, Thatcher WW: Applying nutrition and physiology to improve reproduction in dairy cattle, in Lucy MC, Pate JL, Smith MF, Spencer TE, (eds): *Reproduction in Domestic Ruminants VII*. Nottingham, UK, Nottingham University Press, 2011, pp 387-404.
62. Santos JEP, Narciso CD, Rivera F, Thatcher WW, Chebel RC: Effect of reducing the period of follicle dominance in a timed AI protocol on reproduction of dairy cows. *J Dairy Sci* 93:2976-2988, 2010.
63. Santos JEP, Rutigliano HM, Sá Filho MF: Risk factors for resumption of postpartum cyclicity and embryonic survival in lactating dairy cows. *Anim Reprod Sci* 110:207-221, 2009.
64. Santos JEP, Bilby TR, Thatcher WW, Staples CR, Silvestre FT: Long chain fatty acids of diet as factors influencing reproduction in cattle. *Reprod Dom Anim* 43:23-30, 2008.
65. Santos JE, DePeters EJ, Jardon PW, Huber JT: Effect of prepartum dietary protein level on performance of primigravid and multiparous Holstein dairy cows. *J Dairy Sci* 84:213-224, 2001.
66. Santschi DE, Lefebvre DM, Cue RI, Girard CL, Pellerin D: Complete lactation – milk component yields following a short (35-d) or a conventional (60-d) dry period management strategy in commercial Holstein herds. *J Dairy Sci* 94:2302-2311, 2011.
67. Santschi DE, Lefebvre DM, Cue RI, Girard CL, Pellerin D: Incidence of metabolic disorders and reproductive performance following a short (35-d) or conventional (60-d) dry period management in commercial Holstein herds. *J Dairy Sci* 94:3322-3330, 2011.
68. Sartori R, Haughian JM, Shaver RD, Rosa GJM, Wiltbank MC: Comparison of ovarian function and circulating steroids in estrous cycles of Holstein heifers and lactating cows. *J Dairy Sci* 87:905-920, 2004.
69. Scalia D, Lacetera N, Bernabucci U, Demeyere K, Duchateau L, Burvenich C: In vitro effects of nonesterified fatty acids on bovine neutrophils oxidative burst and viability. *J Dairy Sci* 89:147-154, 2006.
70. Schillo KK: Effects of dietary energy on control of luteinizing hormone secretion in cattle and sheep. *J Anim Sci* 70:1271-1282, 1992.
71. Schneider JE: Energy balance and reproduction. *Physiol Behav* 81:289-317, 2004.
72. Seifi HA, Mohri M, Farzaneh N, Nemati H, Nejhad SV: Effects of anionic salts supplementation on blood pH and mineral status, energy metabolism, reproduction and production in transition dairy cows. *Res Vet Sci* 89:72-77, 2010.
73. von Keyserlingk MA, Olenick D, Weary DM: Acute behavioral effects of regrouping dairy cows. *J Dairy Sci* 91:1011-1016, 2008.
74. Walsh RB, Walton JS, Kelton DF, LeBlanc SJ, Leslie KE, Duffield TF: The effect of subclinical ketosis in early lactation on reproductive performance of postpartum dairy cows. *J Dairy Sci* 90:2788-2796, 2007.
75. Wathes DC, Cheng Z, Chowdhury W, Fenwick MA, Fitzpatrick R, Morris DG, Patton J, Murphy JJ: Negative energy balance alters global gene expression and immune responses in the uterus of postpartum dairy cows. *Physiol Genomics* 39:1-13, 2009.
76. Watters RD, Wiltbank MC, Guenther JN, Brickner AE, Rastani RR, Fricke PM, Grummer RR: Effect of dry period length on reproduction during the subsequent lactation. *J Dairy Sci* 92:3081-3090, 2009.
77. Watters RD, Guenther JN, Brickner AE, Rastani RR, Crump PM, Clark PW, Grummer RR: Effects of dry period length on milk production and health of dairy cattle. *J Dairy Sci* 91:2595-2603, 2008.
78. Wiltbank M, Lopez H, Sartori R, Sangsritavong S, Gümen A: Changes in reproductive physiology of lactating dairy cows due to elevated steroid metabolism. *Therio* 65:17-29, 2006.

SUPERIOR PROTECTION. UNMATCHED ENDURANCE. ONE VACCINE.



**The only viral combination
vaccine to prevent
Lepto hardjo-bovis for
a full 365 days.**

Bovi-Shield GOLD® HB takes disease protection to the next level:

- Helps prevent IBR abortion for 365 days
- Prevents BVD-PI Types 1 and 2 and has a DOI for 365 days
- Prevents Lepto hardjo-bovis infection for 365 days
- Prevents Lepto hardjo-bovis urinary shedding for 365 days
- Single-dose protection — IBR, PI₃, BRSV and BVD Types 1 and 2

Bovi-Shield GOLD HB does it all in one IM or SC dose. It's even backed by a guarantee to prevent BVD-PI calves and IBR abortions.¹ Choose complete reproductive protection from the one you trust.² Talk to your veterinarian or Pfizer Animal Health representative about Bovi-Shield GOLD HB.



**Bovi-Shield
GOLD® HB**

LABEL INDICATIONS: The Bovi-Shield GOLD® line and PregGuard® GOLD FP® 10 are recommended for vaccination of healthy cows and heifers approximately one month prior to breeding. These products also can be administered to pregnant cattle provided they were vaccinated, according to label directions, with any Bovi-Shield GOLD FP or PregGuard GOLD FP vaccine prior to breeding initially and within 12 months thereafter. Failure to follow label directions may result in abortions. The Bovi-Shield GOLD line may be administered to calves nursing pregnant cows, provided their dams were vaccinated within the last 12 months as described above. Consistent with good vaccination practices, heifers should receive at least two vaccine doses, with the second dose administered approximately 30 days prebreeding.

¹This guarantee does not apply to, and Pfizer Animal Health shall not be liable for, any (x) damages caused as a result of the improper handling, misuse or abuse of the vaccines that are the subject of this guarantee, or the willful misconduct or negligence of any third party, or (y) any indirect, punitive, special, incidental or consequential damages. Pfizer Animal Health reserves the right to modify or cancel the terms and conditions of this guarantee.

²Doane Q4 2009.

All brands are the property of Pfizer Inc., its affiliates and/or its licensors. ©2010 Pfizer Inc. All rights reserved. BSD10009

Dairy Wellness Makes a Difference™

