Management Alternatives in Mastitis Control and Eradication Programs

Philip M. Sears, DVM, PhD

Department of Large Animal Clinical Science Michigan State University East Lansing, MI 48824

Herd management programs that are successful in eliminating contagious mastitis include dry cow therapy, segregation and selective culling. When infections involve Streptococcus agalactiae, lactation therapy is still successful although there has been an increase in the number of Strep ag isolates resistant to penicillin and other beta-lactum antibiotics. The major difficulties occur when Staphylococcus aureus is isolated as the primary pathogen in the herd. The primary source of these infections are infected glands and unless these reservoirs are eliminated, new infections will continue to occur. Uninfected cows are mainly exposed during milking but dry cows and prepartum heifers are still at risk when contagious pathogens are present in the lactating herd. Although the method of spread is unknown in nonlactating animals, heifers can become infected before calving with the same strains of organisms as found in the lactating cows.¹ Once Staph aureus infections are eliminated from the lactating animals, the number of infected heifers at calving decreases and often disappears. This is most apparent in herds where Staph aureus has been rapidly eliminated from lactating cows where there is a concurrent rapid decrease in new infections in the heifers at calving. The links speculated as the sources of exposure include insect vectors,² but the reservoir appears to be the adult lactating animal.¹ Because of the spread to young uninfected animals as well as other cows, there is an incentive to try to eliminate these infections from lactating cows. Segregation is an important tool in reducing the risk of exposure at milking, but it does not address other sources of spread. However, many farms cannot create an additional group to segregate infected cows because the group is not the correct size or does not fit well with feeding groups. Likewise, most producers are unwilling to cull infected animals that are producing well and find that the poor response to antibiotic therapy in both lactating and dry cows is frustrating when trying to eliminate Staph aureus from their herds. Since Staph aureus infection can occur anytime during lactation, the longer the lactation and the older the animals, the greater the risk of becoming infected. Thus, herds that have younger cows and a

high turnover rate often have a lower prevalence of *Staph aureus* infected cows, while well established herds with good longevity have greater exposure and accumulate more infections. It is these herds that are trying to retain cows and reach mature production that are often fighting problems with staphylococcal infections. However, no herd is exempt from *Staph aureus*, even with high turnover, especially if they are purchasing animals.

Antibiotic Therapy

Antibiotic treatment during the dry period is still the most effective management practice that we have to eliminate Staph aureus infections. However, even using dry cow therapy, the elimination of Staph aureus seldom exceeds 50% of infections. Treatment during lactation for Staph aureus is often unrewarding because it seldom eliminates the infection.⁴ Antibiotics may lower the number of bacteria and reduce the clinical signs of mastitis, but cows remain infected and continue to have a high somatic cell counts even when the clinical signs are absent. When clinical cases are monitored, many of the clinical cases are observed in the same cows. Thus, once the cow has a clinical case she is more likely to have additional episodes. Milk withdrawal and culling of infected cows are major economic losses on these farms.⁵ Traditional use of intramammary antibiotics during lactation has met with limited success with often less than 10% cures.⁴ In a recent trial measuring efficacy of pirlimycin using intramuscular, intramammary and combination of intramuscular and intramammary treatment, cure rates were only 3.7% for IM, 0% for intramammary and 7.4% for the combination therapy.⁵ However, the same researchers were able achieve a much higher rate of cure (48.8%) using extended therapy of 3 sequential courses of pirlimycin intramammary over 8 days.6 The lack of success with antibiotic therapy has been attributed to inaccessibility, the lack of susceptibility of organisms, limited duration of exposure to antibiotics, and poor immune function. Thus, the response to intramammary antibiotic treatment

is better when the treatment length is extended and the milk SCC is less than 1,000,000 cells/ml.⁷

Role of Vaccination for Staph aureus

Research on immunizing animals against Staph aureus has attempted to identify the best antigens to induce antibody production to prevent new infections. Antigens that have been studied include Protein A, a cell wall component of Staph aureus which interferes with the Fc portion of IgG so that antibody opsonization is inhibited and the organism is not recognized by phagocytic cells and prevents phagocytosis and killing by neutrophils.⁵ Fibronectin, another important virulence factor of Staph aureus, has been used to produce antibody against these surface adhesion factors.⁵ However, some of the most successful immune responses subsequent to vaccination with staphylococcal vaccines occurred when highly encapsulated strains of Staph aureus were used which resulted in a reduction of new infections in heifers after calving.^{9,10} These vaccines have been effective when given to heifers before calving to prevent new infections after challenge in early lactation.¹⁰ Vaccines made using highly encapsulated Staph aureus strains and extractions of exopolysaccharides were effective in reducing new Staph aureus infections at calving from 18.8% in controls to 6.0% in vaccinates, and this protective effect was maintained for at least 6 months.⁵ The study also showed a reduction of clinical signs of mastitis. All of the studies have been aimed at preventing new infections or clinical cases but none of the studies have demonstrated an improvement in eliminating previously existing infections.

Can immune modulation enhance Staph aureus cures?

Since the somatic cell count at time of therapy affects bacterial clearance, it stands to reason that enhancing the immune response could aid in the clearance of bacteria. Although these vaccine preparations have been successful in preventing new infections, vaccination alone has not been successful in eliminating existing infections.⁸ Most bacterins do not produce long lasting antibody levels, but if timed with antibiotic therapy, immune modulation might optimize bacterial clearance.

Traditionally, therapy during lactation has not been considered an economical approach to eliminate *Staph aureus* mastitis and vaccination programs have been aimed at prevention. In a more recent study, we used a vaccine containing both autogenous and capsular antigens to immunize *Staph aureus* infected cows and amplify the cow's immune system against *Staph aureus*, thereby improving the effectiveness of antibiotic therapy. Immune modulation and antibiotic therapy was used in a 48-cow herd on 19 quarters in 10 cows infected with *Staph aureus*. Cows were immunized 2 and 14 days be-

fore starting antibiotic therapy. Extended therapy was used treating each infected quarter with 3 sequential courses of pirlimycin over an 8 day period. A third vaccination was administered during treatment at 7 days into the treatment. All but 3 quarters responded to the antibiotic therapy and all Staph aureus was eliminated by 3 months. The herd bulk tank somatic cell count decreased from 492,000 cells/ml at the time the trial was initiated to 187,000 cells/ml after 3 months and 84,000 one year later. For this herd a calculated cost of \$2656 for identification and treatment of Staph aureus infected cows provided a projected saving of \$5376 to move the DHIA-SCC from a LS 5.3 to LS 2.8 (Table 1). It would only require a 40% cure rate for the saving to equal the expense (cow cure 80% x 49% saving by LS/vaccinationtreatment cost) as calculated from Table 1. However, this does not factor in additional gains by reducing future losses that would have occurred if Staph aureus had remained in the herd.

Table 1.	Economic impact of Staphylococcus aureus	1
elimination	program	

Cost of Pro	Herd				
Herd culture to	50 cows @	\$300.ª			
ID infected cows	\$3.00 twice				
Lactation therapy	24 tubes @ \$3.00	792			
Vaccination	16 Staph	100			
	aureus cows				
Dry cow therapy	5 cows (lact	410			
	+ dry therapy)				
Milk loss for residue	12.00 cwt for	1054			
withdrawal	80 lb/day				
Cull	3 cows @ \$600	1800.ª			
	loss/animal				
Total		\$2656			
Income Savings					
Milk gain based on SCC-LS	LS 5.3 to 2.8	\$5376			
Milk gain based on	Infected vs	\$6846			
ME for Staph aureus	herdmates	+			
Reduced culling for Staph aureus	6 cows @ \$600	\$3600. ^ь			

 $^{\rm a}$ Cost varies with size of herd and animals removed for low economic value

^b Animals removed in 1995 with *Staph aureus* infection for low economic value

Efficacy

In an additional 10 herds tested, the quarter cure rate (Graph 1) following antibiotic therapy ranged from



Graph 1. The distribution (11 herds) of the rate of elimination of *S. aureus* from infected quarters after immune modulation with staphylococcal antigens and six intramammary treatments (3 sequential courses) of pirlimycin hydrochloride.

29% (7 cows) to 95% (30 cows) with a mean herd average cure of 58% for quarters and 54% of cows. There were two groups of cows (14 cows) that received vaccination without antibiotic therapy, in which none of the quarters cleared the Staph aureus infection. These cows were later included into a treatment group and treated with intramammary antibiotic with the same success rate as others in the vaccination-treatment group. There was no difference (P<.001) between cow and quarter rate of cures, although some cows with multiple infected quarters had some quarters clear while other remained infected. Antibiotic treatment was less likely to eliminate all quarter infections in cows when more than 2 quarters were infected. No differences were noted in cows with 1 or 2 infected quarters. Animals in the combination bacterin group responded better to antibiotic therapy, 79% of quarters and 70% of cows, than did animals in the autogenous bacterin group, 36% of quarters 17% of cows (Graph 2). Immune modulation was beneficial since vaccination with both the autogenous or the combination bacterin increased the rate of cures compared to treatment alone.

Discussion

The use of staphylococcal vaccination has been successful in preventing new intramammary infections^{10,11} and reducing the number of clinical cases,¹² however use of a bacterin in infected cows has not affected the persistence of subclinical *Staph aureus* infections. In these studies all vaccinations enhanced the elimination of *Staph aureus* from infected quarters, although there was a difference between the types of antigens (Graph 2). Herd size and number of infected cows and quarters did not affect the overall cure rate as measured for all cows and compared to the mean cure rate of each



Graph 2. The rate of elimination of *S. aureus* following six treatments (3 sequential courses) of pirlimycin hydrochloride after immune modulation with a combination staphylococcal bacterin (experimental), autoenous staphylococcal bacterin or antibiotic treatment only.

herd. The combination bacterin was more effective in eliminating existing infections as measured by quarters and cows. Using cost/benefit analysis Break Even Point (BEP) of 40% based on SCC-LS savings (Table 1), one third (3/11) of the herds (Graph 1) would not have financially benefited from the vaccination-treatment protocol. Only 1 herd exceeded the BEP in the autogenous bacterin group while cows in all herds using the combination bacterin exceeded the BEP (Graph 3).

It is both possible and economically sound (Table 1) to use lactating therapy combined with immune enhancement to develop a mastitis control strategy to eliminate *Staph aureus* from lactating dairy cows. Although selective removal of low value cows and utilizing treatment in the dry period can be an important part of any mastitis control plan, it is not necessary to



Graph 3. Distribution of the rate of elimination of *S. aureus* (by herds) following 3 sequential courses of pirlimycin hydrochloride after immune modulation with a combination staphylococcal bacterin (experimental), autoenous staphylococcal bacterin or antibiotic treatment only. *Break even point* indicates the cure rate level (40%) needed for the economic outcome to equal vaccination and treatment costs.

use these management practices as the sole means of eliminating Staph aureus in the herd.

References

1. Roberson, JR, LK Fox, DD Hancock, JM Gay and TE Besser. 1998 Sources of intramammary infections from Staphylococcus aureus in dairy heifers at first parturition. J Dairy Sci 81:687-694.

2. Owens, WE, SP Oliver, BE Gillispie, CH Ray and SC Nickerson. 1998. Role of horn flies (Haematobia irritans) in Staphylococcus aureus-induced mastitis in dairy heifers. Am J Vet Res, 59:1122.

3. Wilson DJ, PM Sears, et al. 1996. Efficacy of florfenicol for treatment of clinical and subclinical bovine mastitis. Am J Vet Res 57:526. 4. Wilson DJ and JE Lawton. 1999. Dairy cattle identification on cull candidate: Treatment expenses incurred before culling. Agri-Prac 20:16-21

5. Nickerson, SC. 1999. Role of vaccination and treatment programs. In Proc Natl Mastitis Council, Madison, WI, pg 70-85.

6. Owens, WE, CH Ray, RL Bodie and SC Nickerson. 1997. Efficacy of sequential intramammary antibiotic treatment against chronic Staphylococcus aureus intramammary infections. Agri-Pract, 18:10-14.

7. Cattell MB and AP Belschner. 1997. Efficacy of extended lactational intramammary antibiotic treatment against Staphylococcus aureus mastitis. in Proc Natl Mastitis Counc. Albuquerque, NM, p 254.

8. Nickerson SC, WE Owens and RL Boddie. 1993. Effect of a Staphylococc aureus bacterin on serum antibody, new infection, and mammary histology in nonlactating dairy cows. J Dairy Sci. 76:1290. 9. Opdebeeck JP and NL Norcoss . 1983. Frequency and immunologic cross-reactivity of encapsulated Staphylococcus aureus in bovine milk in New York. Am J Vet Res 44(6):96.

10. Sears PM, NL Norcross, et al. 1990. Resistance to Staphylococcus aureus infections in staphylococcal vaccinated heifers. in Proc Int Symp Bovine Mastitis. Natl Mastitis Counc and Am Assoc Bovine Pract, Indianapolis, IN, p 69.

11. Nordhag ML, Nesse LL, Norcross NL and Gudding R. 1994. A field trials with and experimental vaccine against Staphylococcus aureus mastitis in cattle. I. Clinical parameters. J Dairy Sci 77:1276. 12. Yashida K, Ichima Y, Narikow S and Evan WB. 1984. Staphylococcal capsular vaccine for preventing mastitis in two herds in Georgia. J Dairy Sc. 67:620.

Baytril[®] 100

enrofloxacin

100 mg/mL Antimicrobial **Injectable Solution**

For Subcutaneous Use In Cattle Only Not For Use In Cattle Intended For Dairy Production Or In Calves To Be Processed For Veal

BRIEF SUMMARY: Before using Baytril® 100 (enrofloxacin) injectable solution, please consult the product insert, a summary of which follows.

CAUTION: Federal (U.S.A.) law restricts this drug to use by or on the order of a licensed veterinarian. Federal (U.S.A.) law prohibits the extra-label use of this drug in food producing animals.

INDICATIONS:

Baytril® 100 (enrofloxacin) injectable solution is indicated for the treatment of bovine respiratory disease (BRD) associated with *Pasteurella haemolytica*, *Pasteurella multocida* and *Haemophilus somnus*.

DOSAGE ADMINISTRATION:

Single-Dose Therapy: Administer once, a subcuta-neous dose of 7.5-12.5 mg/kg of body weight (3.4-5.7 ml /100 lb)

Multiple-Day Therapy: Administer daily, a subcutaneous dose of 2.5 - 5.0 mg/kg of body weight (1.1-2.3 mL/100 lb). Treatment should be repeated at 24-hour intervals for three days. Additional treatments may be given on days 4 and 5 to animals which have shown clinical improvement but not total recovery.

HUMAN WARNINGS: For use in animals only. Keep out of the reach of children. Avoid contact with eyes. In case of contact, immediately flush eyes with copious amounts of water for 15 minutes. In case of dermal contact, wash skin with soap and water. Consult a physician if irritation persists following ocular or dermal exposures. Individuals with a history of hypersensitivity to quinolones should avoid this product. In humans, there is a risk of user photosensitization within a few hours after excessive exposure to quinolones. If excessive accidental exposure occurs, avoid direct sunlight. To report adverse reactions or to obtain a copy of the Material Safety Data Sheet, call 1-800-633-3796.

> WARNING: Animals intended for human consumption must not be slaughtered within 28 days from the last treatment.

> Do not use in cattle intended for dairy production.

> A withdrawal period has not been established for this product in pre-ruminating calves. Do not use in calves to be processed for yeal

PRECAUTIONS: The effects of enrofloxacin on bovine reproductive performance, pregnancy, and lactation have not been adequately determined.

Subcutaneous injection can cause a transient local tissue reaction that may result in trim loss of edible

Baytril® products. The safety and efficacy of this formulation in species other than cattle have not been determined

Quinolone-class drugs should be used with caution in animals with known or suspected Central Nervous System (CNS) disorders. In such animals, quinolones have, in rare instances, been associated with CNS

stimulation which may lead to convulsive seizures. Quinolone-class drugs have been shown to produce erosions of cartilage of weight-bearing joints and other signs of arthropathy in immature animals of various species. No articular cartilage lesions were observed in the stifle joints of 23-day-old calves at 2 days and 9 days following treatment with enrofloxacin at doses up to 25 mg/kg for 15 consecutive days.

STORAGE CONDITIONS:

Protect from direct sunlight. Do not freeze or store at or above 40° C (104° F).

HOW SUPPLIED:

Baytril® 100 (enrofloxacin) Antimicrobial Injectable Solution:

	Code: 0236	100 mg/mL	100 mL Bottle
	Code: 0321	100 mg/mL	250 mL Bottle
10			

For customer service, to obtain product information, including the Material Safety Data Sheet, or to report adverse reactions call (800) 633-3796. 80002360, R.7

For more information, contact your Bayer representative or call 1-800-633-3796.

