A Review of Bovine Respiratory Disease Vaccine Field Efficacy^a

Louis J. Perino, DVM, PhD Professor West Texas A&M University Breck D. Hunsaker, DVM, MS Manager, Technical Services Schering-Plough Animal Health

Introduction

The ultimate test of a vaccine must be under field conditions and is best obtained from controlled studies of field use.^{1,2,3} Our objective is to review field efficacy of bovine respiratory disease (BRD) vaccines in North American beef cattle, based on research that uses scientifically valid methods with clinically relevant outcomes reported in peer-reviewed publications.

Methods

We conducted a literature search of CAB Abstracts using KnowledgeIndex in CompuServe (Table 1). It included articles from January 1972 to January 1996. We reviewed the 521 citations, deemed 183 relevant to the objective and reviewed them more closely. Additional potentially relevant articles, especially those published before 1972, were identified by less formal means, such as review of bibliographies. There is a debate concerning rigor of the review process between different publications. We did not attempt to differentiate between these. We included all sources that used any review process. We excluded 30 proceedings, transcripts, and abstracts of presentations since they are not routinely peer-reviewed.

 Table 1.
 Literature search strategy.

Search	Criteria*	Citations found
1	bovine or cattle or cow? ^b or calf or calves or bull? or steer? or heifer?	334,972
2	find vaccine? or vaccinat?	31,130
3	find pulmon? or respirat? or pneumon?	58,974
4	(search 1 or search 2) and search 3	521

^aOnly English language articles included.

The materials and methods were reviewed to ensure that the methods were sound, to the degree discernible. Key issues were inclusion of a valid control group, randomization of treatments, blinding of evaluators of subjective outcomes, and adequate statistical power. Methods were also reviewed to ensure that they included clinically relevant outcomes such as morbidity rates, chronic rates, mortality rates, and/or growth performance. Substitution indicators of efficacy, alone, were insufficient to warrant inclusion of the paper in the review. Examples include antibody titer, lymphoproliferation, cytokine levels, and/or safety.

Statistical methods were reviewed to determine appropriateness. When sufficient information was provided, the statistical results were confirmed. If the study met other inclusion criteria but statistical analysis was not done, we attempted to analyze the data.

We were interested in field efficacy, so studies using natural or simulated natural exposure in a field setting were included; studies using experimental challenge models were not. Additionally, reports were excluded when the class of calf, production setting, or vaccine regime used, limited the external validity of the study to beef cattle production in North America.

Results

Based on these criteria, 22 articles remained. Onehundred thirty-seven articles were excluded for one or more of the reasons listed in the methods section. Some difficulties were encountered during the reviews. Optimally, the method of treatment assignment should be clearly and explicitly explained in the materials and methods. This was done in 12 of the 22 reports. This can include formal randomization schemes or systematic schemes that control for important confounding biases, both of which allow for fair evaluation of the treatments. The remainder of the articles (10 of 22) presented statements such as "treatments were assigned randomly". While this suggests that random methods were used, it does not allow the reader to evaluate the validity of the method of randomization. Articles that mentioned randomization were included in the review.

^aAdapted from a paper that appeared in *Optimal Health Management for Enhanced Calf Value*. Proceedings of a Symposium at the 1997 TNAVC, Orlando, FL, 1997. © 1997 Veterinary Learning Systems, Inc.

In some of the unused articles the method of treatment assignment was not mentioned. In this case, it must be assumed that the methods used were, at best, haphazard, and, at worst, biased. For example, some of these articles have severely unbalanced treatment group size, suggesting that a valid treatment assignment scheme was not followed. This results in unreliable conclusions, whether positive or negative, so these articles were excluded.

Determining if evaluators were blinded to treatment assignment was another difficult issue. In many papers, morbidity is the outcome, making unbiased assessment critical to the validity of the results. In 7 of 22 reports, blinding was done and the methods were described in detail. Blinding was not mentioned in 15 of 22 articles. In some designs, it could be inferred that the evaluators would be unlikely to know or be able to determine treatment assignment. In some articles, it was obvious that blinding was not done; in others it was impossible to determine. Articles that did not mention blinding, but met other criteria, were included in the review and footnoted.

Along with blinding of evaluators, a complete and consistently applied case definition is critical to validity of subjective outcomes, such as morbidity. In 5 of 22 papers case definition was not mentioned; however, papers were not excluded on this alone.

Statistics were presented in 18 of 22 reports and statistical methods were discussed in detail in 13 of these. All or some of the authors of 7 of 22 (32%) of the papers were affiliated with the manufacturer or developer of the vaccine reported (Table 2). Nine articles reported positive vaccine effect while 13 reported neutral or negative effects on clinically relevant outcomes.

Table 2. Outcomes of articles and affiliation of authors.

Outcome of trial*	No author affiliation	Author affiliation**	
Positive	3	6	
Neutral	11	1	
Negative	1	0	

*Articles were broadly classified based on the overall results of the study. Neutral or negative results were 2.8-times more likely to be reported in articles with no author affiliation (2-tailed Fishers exact p-value=0.007) **One or all authors worked for or were the manufacturer or developer of the vaccine under study.

Infectious Bovine Rhinotracheitis Virus (IBRV) / Bovine Herpesvirus 1

The IBRV vaccine efficacy studies that met the criteria of this review are equivocal in that results were positive or neutral; however, none were negative. The studies date back to 1958 and 1974 and may not apply to current cattle feeding management practices in North America. In a field trial using modified-live IBRV vaccine at arrival, the incidence of upper respiratory disease was reduced from 17.2% in 3,371 unvaccinated calves to 1% in 3,345 vaccinates (RR=16; p<0.0000).⁴ A well-designed trial using modified-live IBRV5 given at arrival failed to show benefits in health performance. Another report that failed to show IBRV vaccine efficacy involves additional antigens and is presented in the "multiple antigens" section.

Bovine Virus Diarrhea Virus (BVDV)

There are no reliable reports of field trials examining clinical effects of BVDV vaccines in North American beef cattle, based on research that uses scientifically valid methods with clinically relevant outcomes reported in peer-reviewed publications.

Parainfluenzavirus type 3 (PI3V)

Seven field trials which investigated efficacy of vaccination against PI-3 virus infection were done in the 1960's; however, all of these trials had significant design flaws and were excluded from this review.

Bovine Respiratory Syncytial Virus (BRSV)

Mixed results are reported from studies investigating efficacy of BRSV vaccination of calves upon arrival. Statistically significant benefit of BRSV vaccination was shown in auction-market purchased and transported calves with vaccinated calves 2-times less likely to be treated for BRD (OR=2.0, p<0.00001). Freshly weaned and transported calves were 1.4-times less likely to be treated for BRD (OR=1.4; p<.001). Statistically significant benefit of BRSV vaccination was not shown in the two classes of calves with low morbidity rates. These included preconditioned calves (p=0.11) and freshly weaned calves that were not transported (p=0.75).⁶

In a Canadian study (Table 3), results of five separate trials designed to assess BRSV vaccine efficacy were equivocal for calves vaccinated prior to weaning; however, reduction of treatment rate was reported in calves vaccinated once upon arrival. No benefit was found for vaccination upon arrival of yearling cattle.⁷

Two additional trials involving calves^{8,9} and one trial involving stocker cattle¹⁰ all failed to demonstrate benefit of BRSV vaccination at arrival (Table 3).

Pasteurella sp.

Findings reported in the literature are equivocal on the use of more recently available *Pasteurella* sp. vaccines before and at feedlot arrival (Table 4). However, there is a tendency to report only positive results

Table 3.	Summary	of BRSV	vaccine	field	trials.
----------	---------	---------	---------	-------	---------

Vaccine Used & Reference	Calf Type	Vaccination Regime	General Outcome	Specific Outcomes
BRSV-Killed ⁶	Auction-Market purchased and transported	Arrival	Positive	Reduced 60 day treatment rate from 45% to 29% (OR=2.0; p=0.00001)
BRSV-Killed ⁶	Freshly weaned and transported	Arrival	Positive	Reduced 60 day morbidity from 16.5% to 12% (OR=1.4; p=0.001)
BRSV-Killed ⁶	Preconditioned	30 days prior to shipment	Neutral	3.4% vx vs. 2.2% ctrl treatment rate (p=0.11)
BRSV-Killed ⁶	Freshly weaned and non-transported	Arrival	Neutral	1.3% vx vs. 0.5% ctrl treatment rate (p=0.75)
BRSV-MLV7	138 bull calves sent to bull test station at weaning	3 weeks prior to weaning & boostered at weaning	Positive	120 day morbidity rate reduced from 17% to 5%. (p<.0.05)
BRSV-MLV ⁷	97 bull calves sent to custom feedlot in southern Alberta	3 weeks prior to weaning & boostered at weaning	Neutral	
BRSV-MLV ⁷	317 heifer calves and 52 bull calves remaining at ranch	3 weeks prior to weaning & boostered at weaning	Neutral	
BRSV-MLV ⁷	283 bull calves arriving at bull test station	Arrival	Neutral	
BRSV-MLV ⁷	253 Charolais-cross calves at research station	Weaning and boostered 3 weeks later	Neutral (adg)	
BRSV-MLV7	611 market-derived yearlings sent to commercial feedlot	Arrival	Neutral	
BRSV-MLV ⁷	4913 yearlings & 1716 calves in large commercial feedlot	Arrival	Positive for calves; Neutral for yearlings	21% treatment rate in ctrl vs. 17% treatment rate in vx's; 8 wk observation period (p<0.05)
BRSV-MLV ⁸	422 calves of undefined wt, age, breed	Arrival	Neutral	
BRSV- indefined ⁹	192 calves; 96 in 1984 & 96 in 1985	Arrival	Neutral	
BRSV- indefined ¹⁰	754 stocker cattle	Arrival	Neutral	

Table 4. Summary of Pasteurella vaccine field trials.

Vaccine Used	Calf Type	Vaccination	General	Specific Outcomes
P. haemolytica toxoid ¹¹	560 to 827 lb. calves received in Oct. to Dec. in Canada	Arrival	Positive	Reduced 64-124 day mortality from 4.2% to 2.2%
P. haemolytica toxoid ¹²	Spring-born, Hereford-cross ranch calves in southwest TX	Branding & arrival or arrival only	Positive	Reduced 60 day morbidity from 31% to approx. 20% & mortality from 2.5% to approx. 0.2%
P. haemolytica toxoid ¹³	300 lb. calves hauled from KY to NM	Arrival & 14 days post-arrival	Positive	Reduced 28 day morbidity from 50% to 13% and mortality from 6.7% to 0%
P. haemolytica toxoid ¹⁴	Auction market calves hauled from southeast US to OK	Preshipment or postshipment (arrival)	Neutral	
P. haemolytica toxoid ¹⁵	500 to 600 lb., 6 to 8 mo. Old ranch calves in Canada	3 weeks before shipment &/or shipment	Neutral	
Streptomycin- dependent P. multocida & haemolytica ¹⁶	300 to 600 lb. Preconditioned calves	14 days before shipment &/or arrival	Positive	Increase gain from 0.83 kg/day to approx. 1.1 kg/day
Streptomycin- dependent P. multocida & haemolytica ¹⁶	410 to 600 lb. Auction market calves	Arrival	Neutral	
Live intradermal P. haemolytica ¹⁷	534 lb. Auction market calves from southeast US	Arrival	Positive	Reduced 28 day morbidity from 40% to 33% and mortality from 1.2% to 0%.
Tissue-culture derived P. haemolytica bacterin ¹⁹	Weaned calves hauled from TN to order-buyer barn, then to TX	98 to 95 days before weaning and arrival (8 days after weaning)	Neutral	
P. haemolytica capsular antigen	290 lb. calves hauled ¹⁸ from FL to OK	Arrival	Neutral	

in the scientific literature; thus, negative or neutral findings remain unpublished. This bias, along with the fact that there are reports that fail to show a positive effect, raises doubt concerning efficacy of *Pasteurella* sp. vaccines when used in the field.

The largest body of *Pasteurella* vaccine data exists for *P. haemolytica* toxoid. Three studies have shown statistically significant reduction in morbidity and/or mortality in calves administered a *P. haemolytica* toxoid at arrival.^{11,12,13} However, two clinical trials showed no significant effects when the same vaccine was given at arrival¹⁴ or three weeks before shipment and/or arrival.¹⁵ In no reports was health performance in vaccinates negatively affected.

There are individual reports on various other commercial or experimental *Pasteurella* sp. vaccines. These include reports of significant efficacy in field studies of a streptomycin-dependent live *Pasteurella* sp. vaccine¹⁶ and an intradermally-administered live *P. haemolytica* vaccine.¹⁷ Alternatively, a field study of a *P. haemolytica* capsular antigen vaccine failed to show significant health effects,¹⁸ as did a study using a tissue-culturederived *P. haemolytica* bacterin.¹⁹

For some currently available *Pasteurella* sp. vaccines there are no reports meeting the objectives and criteria of this review. There are reports of lack of field efficacy with earlier *Pasteurella* sp. bacterins.^{20,21} There is also a report of increased health problems following vaccination with earlier *Pasteurella* sp. bacterins.²² However, this study did not mention if treatment assignment was randomly done and the experimental unit is unclear, making the validity of the data analysis suspect.

Haemophilus somnus

As with other vaccine antigens of bovine respiratory disease prophylaxis, results of field trials evaluating efficacy of H. somnus bacterins have been conflicting (Table 5). A 1984 review of this subject concluded that this is the case.¹ One group of investigators has reported negative effects of vaccination once with a commercial H. somnus bacterin in that significantly more animals in groups of calves vaccinated once were treated for respiratory disease as compared to groups of unvaccinated control calves or groups of calves vaccinated twice at a 21 day interval.²³ However, these findings conflicted with earlier reports by these authors that no significant difference in the number of animals treated was found between groups of calves immunized once with a commercial H. somnus bacterin and groups of unimmunized control calves.²¹ Conversely, these investigators had reported earlier that morbidity (number of animals treated for respiratory disease) was significantly reduced in groups of calves vaccinated with a commercial H. somnus bacterin upon arrival at the feedlot and re-vaccinated 21 days later as compared to groups vaccinated twice with a bivalent P. haemolytica, P. multocida bacterin or unvaccinated controls.²⁰

Table 5. Summary of Haemophilus somnus vaccinefield trials.

Vaccine Used & Reference	Calf Type	Vaccination Regime	General Outcome	Specific Outcomes
H. somnus bacterin ²³	20 crossbred steers	3 groups: 1. unvaccinated 2. vaccinated at arrival 3. vaccinated at arrival & boostered 21 days later	Neutral to negative	grp 2 had significantly higher treatment rate than grps 1 & 3
H. somnus bacterin ²¹	306 crossbred steers	Arrival	Neutral	
H. somnus bacterin ²⁰	340 crossbred heifers	Arrival & boostered	Positive 21 days later	reduced treatment rate from 21/106 vx vs. 33/ 107 ctrl

Mycoplasma sp.

While there are some reports on *Mycoplasma* sp. vaccines, there are no reliable reports of field trials evaluating efficacy of vaccines in North American beef cattle, based on research that examines clinically relevant outcomes with scientifically valid methods in peer-reviewed publications.

Coronavirus, Chlamydia, Adenovirus, Calicivirus

There are no reports of vaccine field trials for these pathogens, germane to North American beef-cattle production in peer-reviewed publications.

Multiple antigens

Field trials were carried out with vaccinates receiving multiple antigens, making it impossible to determine the effects of individual antigens. These can be subdivided into two broad groups: vaccine administered at or near the time of feedlot arrival and vaccine administered several weeks before feedlot arrival.

Assuming valid design, execution, and analysis, interpretation of the first group is fairly straightforward. Some studies of arrival vaccination suggest it does not affect or may even decrease health performance. A well designed study using modified-live IBRV and PI3V vaccine along with a *Pasteurella haemolytica* toxoid failed to show health performance benefits.²⁴ This is supported by findings in a multi-year observational study in Ontario, Canada, which reported that administration of respiratory vaccines (IBRV or IBRV-PI3V or IBRV-PI3V-*Pasteurella*) to calves vaccinated within two weeks of arrival was associated with increased risk of mortality (relative risk = 2.4).²⁵ In contrast, subcutaneous vaccination with a *P. haemolytica* and *H. somnus* vaccine at arrival reduced BRD morbidity from 41% to 29%.²⁶

The second type of mixed-antigen study is when vaccines are administered several weeks before feedlot arrival. These are often part of a preconditioning or preweaning study. Since an unvaccinated, but similarly managed group, is rarely included in these studies, the effects of management interventions such as preweaning and bunk acclimation are totally confounded with vaccine effect. It is impossible to know which accounts for improvements in health performance.

Controlled studies of these systems have routinely shown improved health performance. When control groups of calves experience low morbidity rates, no preconditioning effect can be demonstrated. Because it is not possible to separate the effects of vaccination from other management interventions, these papers were not included in this review; however, review papers of preconditioning have been published.²⁷

Conclusion

We are impressed with the small number of useful reports of clinical efficacy in field settings for BRD vaccines. While there are hundreds of reports in the literature, most suffer from one or more design flaws or limitations. Many of these are referenced in support of vaccination. In particular, published data supporting BRD vaccination at arrival in North American feedlots is equivocal, at best.

Since licensing of BRD vaccines is based, in part, on demonstration of efficacy and these vaccines are used widely in North American beef cattle production, it appears that this is largely based on biologically logical extrapolation from challenge studies, rather than field efficacy data. Controlled laboratory evaluations are an important and necessary step in vaccine development. They not only provide evidence supporting efficacy, but also frequently give invaluable insight into mechanisms of protective immunity. Studies of immune mechanisms are critical to our understanding of host-pathogen interactions and how to induce protective immunity. This insight is the basis for much of the progress in vaccine improvement. However, demonstration of activation of these immune mechanisms is not *de facto* proof of clinical efficacy in a field setting. Laboratory experiments are not substitutes for field trials and extrapolation of findings from the laboratory to the field is fraught with pitfalls.

As mentioned at the outset, the ultimate test of a vaccine must be under field conditions and is best obtained from controlled studies of field use. Economic justification for vaccination is based on this information, compared to vaccination costs. Each producer will place a different value on vaccine attributable benefits, such as improved health and/or growth performance. These values will also change with fluctuating market values of cattle and feed. This makes determining the cost:benefit ratio of vaccination a moving target. However, addressing this issue requires the clinically relevant, statistically significant differences we tried to identify in this review. Without sound field trial design and execution, which ensures the information is reliable, and statistical significance, which ensures the differences are real, clinical outcomes cannot be extrapolated to economic justification.

It seems prudent to mention that we do not believe that this review suggests that we should abandon vaccination of cattle for BRD before or at arrival. Rather, it suggests that we may be making less than optimal recommendations on vaccine use because of a lack of clinically relevant information. We believe this creates an opportunity.

It is time to critically evaluate vaccination as a management tool. Since it has not been done by the government, universities, or manufacturers yet, it seems unlikely that it will be done very soon unless we take the initiative. There already exists a number of unusable studies, we suggest that those interested in undertaking this challenge be uncompromising in their experimental design. To be reliable, studies must:

- · include a valid control group,
- \cdot use an externally relevant population,
- use a clinically reasonable treatment regime,
- use random treatment assignment,
- · blind assessors to treatment assignment,
- use a field challenge in an externally relevant production setting,
- · ensure adequate follow-up,
- have adequate statistical power to detect treatment effects,
- · control for confounding variables,
- use appropriate statistical evaluation to determine if differences are real,
- measure clinically relevant outcomes to determine if differences are important.

All the above should be described in detail in the methods and, along with the results, be published in a peer-reviewed source. Reviewers should include these items in criteria used to evaluate submissions. Reports published in peer-reviewed journals are subject to formal scientific scrutiny and are available to the entire veterinary medical community. Access to reliable, clinically relevant information will enhance practical decision making.

References and reason(s) for exclusion.

1. Martin SW. Vaccine prophylaxis of bovine respiratory disease. Can Vet J 1984;25:44-48.^k 2. Wilkie BN. Is immunization against bovine respiratory disease possible? Can Vet J 1984;25:48-50.k 3. Confer AW, Panciera RJ, Mosier DA. Bovine pneumonic pasteurellosis: immunity to Pasteurella haemolytica. JAm Vet Med Assn 1988;193:1308-1316.^k 4. York CJ, Schwarz AJF, Zirbel L. et al. Infectious bovine rhinotracheitis vaccine. Vet Med 1958;Oct:522-524.^a 5. Curtis RA, Angulo A. A field trial to evaluate an intranasal infectious bovine rhinotracheitis vaccine. Can Vet J 1974;15:327-330.ª 6. Hansen DE, Syvrud R, Armstrong D. Effectiveness of a Bovine Respiratory Syncytial Virus Vaccine in Reducing the Risk of Respiratory Disease. Agri-Practice 1992;13:19-22. 7. Van Donkersgoed J, Janzen ED, Townsend HG, Durham PJ. Five field trials on the efficacy of a bovine respiratory syncytial virus vaccine. Can Vet J 1990;31:93-100. 8. Baker JC, Rust SR, Ciszewski DK, Coe PH. A safety trial of a bovine respiratory syncytial virus vaccine in feedlot calves. Bov Practitioner 1986;21:70-72.ª 9. Morter RL, Amstutz HE. Effectiveness of vaccination of feedlot cattle with bovine respiratory syncytial virus (BRSV). Bov Practitioner 1986;21:67-69.ª 10. Johnson BD, Hays VS, Gill DR, Smith RA, Owens FN, Ball RL. Respiratory syncytial virus vaccine for stressed stocker cattle. Oklahoma Agricultural Experiment Station Animal Science Research Report 1988; MP-125:105-110.ª 11. Jim K, Guichon T, Shaw G. Protecting feedlot calves from pneumonic pasteurellosis. Vet Med 1988;83:1084-1087.ª 12. Bechtol DT, Ballinger RT, Sharp AJ. Field trial of a Pasteurella haemolytica toxoid administered at spring branding and in the feedlot. Agri-Practice 1991;12:6-14. 13. Macolm-Callis KJ, Galyean ML, Duff GC. Effects of dietary supplemental protein source and a Pasteurella haemolytica toxoid on

performance and health of newly received calves. Agri-Practice 1994;15:22-28.ª 14. McLean GS, Smith RA, Gill DR, et al. An evaluation of an inactivated, leukotoxin-rich, cell-free Pasteurella hemolytica vaccine for prevention of undifferentiated bovine respiratory disease. Oklahoma State University Animal Science Research Report 1990;MP-129:135-140.ª 15. Thorlakson B, Martin W, Peters D. A field trial to evaluate the efficacy of a commercial Pasteurella haemolytica bacterial extract in preventing bovine respiratory disease. Can Vet J 1990;31:573-579. 16. Kadel WL, Chengappa MM, Herren CE. Field-trial evaluation of a Pasteurella vaccine in preconditioned and nonpreconditioned lightweight calves. Am J Vet Res 1985;46:1944-1948.^a 17. Smith RA, Gill DR, Hicks RB. Improving the performance of stocker and feedlot calves with a live Pasteurella haemolytica vaccine. Vet Med 1986;81:978-981.ª 18. Hill WJ, Kirkpatrick J, Gill DR, et al. The effects of Septimune on health and performance of stressed stocker cattle. Oklahoma State University Animal Science Research Report 1993; P-933:301-303.ª 19. Frank GH, Briggs RE, Loan RW, et al. Respiratory tract disease and mucosal colonization by Pasteurella haemolytica in transported calves. Am J Vet Res 1996;57:1317-1320.ª 20. Amstutz HE, Horstman LA, Morter RL. Clinical evaluation of the efficacy of Haemophilus somnus and Pasterrella sp. bacterins. Bov Practitioner 1981;16:106-108.ª 21. Morter RL, Amstutz HE, Crandell RA. Clinical evaluation of prophylactic regimens for bovine respiratory disease. Bov Practitioner 1982;17:56-58.ª 22. Bennett BW. Efficacy of Pasteurella bacterins for yearling feedlot cattle. Bov Practice 1982;3:26-30.^{a,b,f} 23. Morter RL, Amstutz HE. Evaluating the efficacy of a Haemophilus

somnus bacterin in a controlled field trial. Bov Practitioner 1983;18:82-83.* 24. Bateman KG. Efficacy of a Pasteurella haemolytica vaccine/ bacterial extract in the prevention of bovine respiratory disease in recently shipped feedlot calves. Can Vet J 1988;29:838-839. 25. Martin SW, Meek AH, Davis DG, et al. Factors associated with mortality and treatment costs in feedlot calves: the Bruce County Beef Project, years 1978, 1979, 1980. Can J Comp Med 1982;46:341-349. 26. Van Donkersgoed J, Schumann FJ, Harland RJ, et al. The effect of route and dosage of immunization on the serological response to a Pasteurella haemolytica and Haemophilus somnus vaccine in feedlot calves. Can Vet J 1993;34:731-735. 27. Cole NA. Preconditioning calves for the feedlot. Vet Clin North Am 1985;1:401-412.^k

References not used.

28. Baker JC, Velicer LF. Bovine respiratory syncytial virus vaccination: current status and future vaccine development. Comp Cont Ed Practicing Veterinarian 1991;13:1323-1334.^k 29. Bartha A. Immunization of cattle with a polyvalent bovine Adenovirus vaccine. Dev Biol Stan 1974;26:15-18.ⁱ 30. Berneri C, Amadori M, Ceccarelli A, et al. Comparative evaluation of specific vaccines and immuno-modulators in disease control of beef cattle. J Vet Med. B 1991;38:60-77.g,ⁱ 31. Bohlender RE. Field trials of a bovine respiratory syncytial virus vaccine. Mod Vet Practice 1984;65:606-609.a,b 32. Cardella MA, Adviento MA, Nervig RM. Vaccination studies against experimental bovine Pasteurella pneumonia. Can J Vet Res 1987;51:204-211.^h 33. Carmel DK, Barao SM, Douglass LW. Effects of vaccination against 18 immunogens in beef replacement heifers at weaning. J Am Vet Med Assn 1992;201:587-590.i 34. Catt DM, Chengappa MM, Kadel WL, et al. Preliminary studies with a live streptomycin-dependent Pasteurella multocida and Pasteurella haemolytica vaccine for the prevention of bovine pneumonic pasteurellosis. Can J Comp Med 1985;49:366-371.h 35. Chengappa MM, McLaughlin BG, Kadel WL, et al. Bovine pneumonic pasteurellosis: efficacy testing a live vaccine. Vet Med 1988;83 (8:837-840.^h 36. Chengappa MM, McLaughlin BG, Kadel WL, et al. Efficacy of a live Pasteurella multocida vaccine for the prevention of experimentally induced bovine pneumonic pasteurellosis. Vet Micro 1989;21:147-154.h 37. Confer AW, Panciera RJ, Corstvet RE, et al. Bovine pneumonic pasteurellosis: effect of culture age of Pasteurella haemolytica used as a live vaccine. Am J Vet Res 1984; 45:2543-2545.h

38. Confer AW, Panciera RJ, Fulton RW, et al. Effect of vaccination with live or killed Pasteurella haemolytica on resistance to experimental bovine pneumonic pasteurellosis. Am J Vet Res 1985; 46:342-347.^h 39. Confer AW, Panciera RJ, Gentry MJ, et al. Immunologic response and resistance to experimentally induced pneumonic pasteurellosis in cattle vaccinated with various dosages of lyophilized Pasteurella haemolytica. Am J Vet Res 1986; 47:1853-1857^h 40. Confer AW, Panciera RJ, Gentry MJ, et al. Immunologic response to Pasteurella haemolytica and resistance against experimental bovine pneumonic pasteurellosis, induced by bacterins in oil adjuvants. Am J Vet Res 1987; 48:163-168.^h 41. Confer AW, Panciera RJ, Mosier DA. Bovine pneumonic pasteurellosis: immunity to Pasteurella haemolytica. J Am Vet Med Assn 1988; 193:1308-1316.^k 42. Confer AW, Simons KR, Panciera RJ, et al. Serum antibody response to carbohydrate antigens of Pasteurella haemolytica serotype 1: relation to experimentally induced bovine pneumonic pasteurellosis. Am J Vet Res 1989; 50:98-105.h 43. Confer AW, Panciera RJ. Testing of two new generation Pasteurella haemolytica vaccines against experimental bovine pneumonic pasteurellosis. Agri-Practice 1994; 158:10-15.h 44. Conlon JA, Shewen PE, Lo RYC. Efficacy of recombinant leukotoxin in protection against pneumonic challenge with live Pasteurella haemolytica A1. Infect and Immunity 1991; 59:587-591.^h 45. Conlon JA, Shewen PE, Lo RYC. Efficacy of recombinant leukotoxin in protection against pneumonic challenge with live Pasteurella haemolytica A1. Infect and Immunity 1991; 59:587-591.^h 46. Conlon JAR, Gallo GF, Shewen PE, Adlam C. Comparison of protection of experimentally challenged cattle vaccinated once or twice with a Pasteurella haemolytica bacterial extract vaccine. Can J Vet Res 1995; 59:179-182.h 47. Cravens RL. Clinical response of feeder calves under direct IBR and BVD virus challenge: a comparison of two vaccines and negative control. Bov Practitioner 1991; 26:154-158.h 48. Darcel Cle Q, Jericho KWF. Failure of a subunit bovine herpesvirus 1 vaccine to protect against experimental respiratory disease in calves. Can J Comp Med 1981; 45:87-91.^h 49. Ellis JA, Davis WC, Talens L, et al. Clinical and immunologic response of cattle to administration of a vaccine containing modified-live bovine respiratory syncytial virus. J Am Vet Med Assn 1990; 196:583-589.ⁱ 50. Fulton RW. Confer AW, Burge LJ, et al. Antibody responses by cattle after vaccination with commercial viral vaccines containing bovine herpesvirsu-1, bovine viral diarrhea virus, parainfluenza-3 virus, and bovine respiratory syncytial virus immunogens and subsequent revaccination at day 140. Vaccine 1995; 13:725-733.ⁱ 51. Frankena K, Klaassen C HL, Bosch JC, et al. Double blind field evaluation of a trivalent vaccine against respiratory disease in veal calves. Vet Quarterly 1994; 163:148-152.^g 52. Friend SCE, Wilkie BN, Thomson RG, et al. Bovine pneumonic pasteurellosis: experimental induction in vaccinated and nonvaccinated calves. Can J Comp Med 1977; 41:77-83.^h 53. Frerichs GN, Woods SB, Lucas MH, et al. Safety and efficacy of live and inactivated infectious bovine rhinotracheitis vaccines. Vet Record 1982; 111:116-122.h.i 54. Gentry MJ, Confer AW, Panciera RJ. Serum neutralization of cytotoxin from Pasteurella haemolytica, serotype 1 and resistance experimental bovine pneumonic pasteurellosis. Vet Immun and Immunopath 1985; 9:239-250.h 55. Gilmour NJL, Angus KW, Donachie W, et al. Vaccination against experimental pneumonic pasteurellosis. Vet Record 1982; 110:450.h 56. Gilmour NJL, Gilmour JS, Donachie W, et al. Failure of a Pasteurella haemolytica extract vaccine to protect calves against experimental pneumonic pasteurellosis. Vet Record 1987; 121:277-278.h 57. Groom SC, Little PB. Effects of vaccination of calves against induced Haemophilus somnus pneumonia. Am J Vet Res 1988; 49:793-800.^{a,b,h,i} 58. Gutekunst DE, Paton IM, Volenec FJ. Parainfluenza-3 vaccine in cattle: comparative efficacy of intranasal and intramuscular routes. JAm Vet Med Assn 1969; 155:1879-1885. a.b.e 59. Hall RF, Williams JM, Smith GL. Field evaluation of Haemophilus somnus bacterin. Vet Med Sm An Clin 1977; Sept:1368-1370.ⁱ 60. Harland R J, Potter AA, van Drunen Littel-van den Hurk S, et al. The effect of subunit or modified live bovine herpesvirus-1 vaccines on the efficacy of a recombinant Pasteurella haemolytica vaccine for the prevention of respiratory disease in feedlot calves. Can

Vet J 1992;33:734-741.º 61. Hjerpe CA. Bovine vaccines and herd vaccination programs. Vet Clin N Am Food An Practice 1990; 6:167-260.* 62. Howard CJ, Gourlay RN, Taylor G. Induction of immunity in calves to Mycoplasma bovis infection of the respiratory tract. Vet Micro1 977;2:29-37.g.h 63. Howard CJ, Gourlay RN, Taylor G. Immunity to Mycoplasma bovis infections of the respiratory tract of calves. Res Vet Sci 1980; 28:242-249.g,i 64. Howard CJ, Gourlay RN, Taylor G. Immunity to Mycoplasma infections of the calf respiratory tract. Adv Exp Med Biol 1981; 137:711-726.k 65. Howard CJ, Stott EJ, Thomas LH, et al. Protection against respiratory disease in calves induced by vaccines containing respiratory syncytial virus, parainfluenza type 3 virus, Mycoplasma bovis and M. dispar. Vet Record 1987; 121:372-376.^{fg} 66. Howard CJ, Stott JE, Thomas LH, et al. Vaccination against natural outbreaks of respiratory disease in calves associated with Mycoplasma bovis, M. dispar, and respiratory syncytial virus infection. Zbl Suppl 1990;20:401-405.g 67. Howard CJ, Clarke MC, Sopp P, et al. Systemic vaccination with inactivated bovine virus diarrhoea virus protects against respiratory challenge. Vet Micro 1994; 42:171-179.h 68. Jericho KWF, Magwood SE, Stockdale PHG. Prevention of experimental bovine pneumonic pasteurellosis by exposure to IBR virus. Can Vet J 1976; 17:194-195.h 69. Jericho KWF, Langford EV. Aerosol vaccination of calves with Pasteurella haemolytica against experimental respiratory disease. Can J Comp Med 1982; 46:287-292.h 70. Jericho KWF, Yates WDG, Babiuk LA. Bovine herpesvirus-1 vaccination against experimental bovine herpesvirus-1 and Pasteurella haemolytica respiratory tract infection: onset of protection. Am J Vet Res 1982; 43:1776-1780.h 71. Jericho KWF, Babiuk LA. The effect of dose, route and virulence of bovine herpesvirus 1 vaccine on experimental respiratory disease in cattle. Can J Comp Med 1983; 47:133-139.h 72. Jericho KWF, Cho HJ, Kozub GC. Protective effect of inactivated Pasteurella haemolytica bacterin challenged in bovine herpesvirus-1 experimentally infected calves. Vaccine 1990; 8:315-320.h 73. Jericho KWF, Loewen KG, Smithson SE, et al. Protective effect of inactivated bovine herpesvirus-1 in calves experimentally infected with bovine herpesvirus-1 and Pasteurella haemolytica. Res in Vet Sci 1991; 51:209-214.h 74. Kahrs RF. Rational basis for an immunization program against the common diseases of the bovine respiratory tract. Can Vet J 1974; 15:252-256.^k 75. Kahrs RF. Infectious bovine rhinotracheitis: a reupdate. J Am Vet Med Assn view and 1977: 171:1055-1064.^k 76. Karst, O, Mitchell, S. Intranasal vaccination of cattle with an attenuated Gladysdale strain of Mycoplasma mycoides var. mycoides. J Comp Path 1972; 82:171-178. a,b.g,m 77. Kelling, CL. Controlling BRSV (bovine respiratory syncytial virus) infection in calves. Vet Med 1993; 88:903-906.k 78. Kennedy JA. The effects of Re-17 mutant Salmonella typhimurium bacterin-toxoid on bovine respiratory disease in feedlot heifers. Agri-Practice 1995;16:29-31. a.d.f. 79. Kimman TG, Sol J, Westenbrink F, et al. A severe outbreak of respiratory tract disease associated with bovine respiratory syncytial virus probably enhanced by vaccination with modified live vaccine. Vet Quarterly 1989; 11:250-253. e.g 80. King NB, Gale C. Studies on myxovirus parainfluenza-3 vaccine for prevention of shipping fever in cattle. J Am Vet Med Assn 1963; 142:881-883.a.b.c 81. Kita J, Oyrzanowska J, Prandota J. Evaluation of the attenuated vaccine Para-Ribovac against bovine parainfluenza and IBR. Zbl Vet Med B 1983; 30:502-511.g.h.i 82. Kolar JR, Shechmeister IL, Kammlade WG. Use in cattle of formalin-killed polyvalent vaccine with adjuvant against infectious bovine rhinotracheitis, bovine viral diarrhea, and parainfluenza-3 viruses. AmJVet Res 1972: 33:1415-1420.ⁱ 83. Kubota M, Fukuyama S, Takamura K, et al. Field trials on a live bovine respiratory syncytial virus vaccine in calves. J Vet Medical Sci 1992; 54:957-962.^g 84. Kucera CJ, Wong JCS, Feldner TJ. Challenge exposure of cattle vaccinated with a chemically altered strain of Pasteurella haemolytica. Am J Vet Res 1983;44:1848-1852.^h 85. Kucera CJ, Feldner TJ, Wong JCS. The testing of an experimental bovine respiratory syncytial virus vaccine. Vet Med Sm An Clinician 1983; 78:1599-1604.^{a,b} 86. Loan RW, Tigges MG, Purdy CW. A tissue culture-derived Pasteurella haemolytica vac-

cine. Bov Practitioner 1989;24: 22-24. a.b 87. Lopez JW, Woods GT, Crandell RA, et al. A preconditioning program for beef calves. Agri-Practice 1984; 5:7-19.º 88. Mann DD, Buening GM, Thorne JG. Efficacy of aerosol, intranasal and intramuscular vaccination against selected bovine viral diseases. Cornell Veterinarian 1983; 73:375-379.^h 89. Mansfield ME, Crandell RA, Woods GT. The effect of intranasal IBR-PI3 vaccination of feeder cattle on arrival at a feedlot compared with vaccination at the start of an outbreak of acute respiratory tract disease. Bov Practitioner 1978; 13:83-87.b Marshall, RG. Antibody response of calves after intranasal inoculation with parainfluenza-3 virus and resistance of inoculated calves to experimental homologous viral infection. Am J Vet Res 1981; 42:907-911.^h 91. Martin SW. Vaccination: is it effective in preventing respiratory disease or influencing weight gains in feedlot calves? Can Vet J 1983;24:10-19.^k 92. Martin W, Willson P, Curtis R, Allen B, Acres S. A field trial of preshipment vaccination with intranasal infectious bovine rhinotracheitis-parainfluenza-3 vaccines. Can J vine Comp Med 1983;47:245-249.^j 93. Martin SW. Vaccine prophylaxis of bovine respiratory disease. Can Vet J 1984; 25:44-48.^k 94. Martin W, Acres S, Janzen E, et al. A field trial of preshipment vaccination of calves. Can Vet J 1984; 25:145-147.¹ 95. Matsumoto M, Schmitz JA, Syuto B, et al. Immunogenicity of a soluble antigen against Pasteurella haemolytica-associated pneumonia in calves. Vet Res Communications 1984; 8:117-130.^h 96. Mattson DE, Wangelin JR, Sweat RL. Vaccination of dairy calves with bovine adenovirus type 3. Cornell Veterinarian 1987; 77:351-361.^g 97. McKercher DG, Crenshaw GL. Comparative efficacy of intranasally and parenterally administered infectious bovine rhinotracheitis vaccines. J Am Vet Med Assn 1971;159:1362-1369.^h 98. McKercher DG. Nasal versus parenteral vaccination for the protection of cattle against viral infection of the respiratory tract. Arch Vet Ital 1972; 23:123-134.h, 99. Mee JF, Wafa S, O'Farrell KJ. Clinical response and economic value of vaccinating dairy calves with two types of vaccines (bovine respiratory syncytial virus and bovine parainfluenza virus-3. Agri-Practice 1995; 16:15-19.^g 100. Mohanty SB, Lillie MG, Ingling AL. Effect of serum and nasal neutralizing antibodies on bovine respiratory syncytial virus infection in calves. J Inf Dis 1976;134:409-413.^h 101. Mohanty S B, Rockemann DD, Davidson JP, et al. Effect of vaccinal serum antibodies on bovine respiratory syncytial viral infection in calves. Am J Vet Res 1981; 42:881-883.h 102. Morzaria SP, Richards MS, Harkness JW, et al. A field trial with a multicomponent inactivated Record viral vaccine. Vet 1979; 105:410-414^{.a,c,g} 103. Newman PR, Corstvet RE, Panciera RJ. Distribution of Pasteurella haemolytica and Pasteurella multocida in the bovine lung following vaccination and challenge exposure as an indicator of lung resistance. Am J Vet Res 1982; 43:417-422.^h 104. Oliphant, R. Effect of Re-17 mutant Salmonella typhimurium bacterin-toxoid on respiratory disease and production. Agri-Practice 1994; 15:13-16.^d 105. Panciera RJ, Corstvet RE, Confer AW, et al. Bovine pneumonic pasteurellosis: effect of vaccination piratory with live Pasteurella species. Am J Vet Res 1984; 45:2538-2542.^h 106. Phillip J IH, Clegg FG, Halliday GJ, et al. An examination of two bovine respiratory disease vaccines. Vet Record 1973;92:420-424.g,ij 107. Phillips RM, Heuschele WP, Todd JD. Evaluation of a bovine viral diarrhea vaccine produced in a porcine kidney cell line. Am J Vet Res 1975;36:135-140.e,h 108. Pollreisz JP, Jordan T. Preshipment programs for beef cattle. Comp Cont Ed Practicing Veterinarian 1988;10:367-371.k 109. Primal SV, Silva S, Little PB. The protective effect of vaccination against experimental pneumonia in cattle with Haemophilus somnus outer membrane antigens and interference by lipopolysaccharide. Can J Vet Res 1990;54:326-330.h 110. Probert M, Stott EJ, Thomas LH, et al. An inactivated parainfluenza virus type 3 vaccine: the influence of vaccination regime on the response of calves and their subsequent to challenge. Res Vet Sci 1978; 24:222-227.^{a,b,c} 111. Radostits OM. The control of infectious diseases of the respiratory and digestive tracts of cattle. Can Vet J 1991; 32:85-89.^k 112. Ribble CS, Jim GK, Janzen ED. Efficacy of Immuni-

bacterin. Can J Vet Res 1988; 52:191-198.^f 113. Ribble CS. Assessing vaccine efficacy. Can Vet J 1990:31:679-681.^k 114. Schell K. Sanderson RP, Whalen JW, et al. The antigenicity of multivalent vaccines for bovine respiratory disease. Cornell Veterinarian 1972; 62:101-109.ⁱ 115. Schultz RH, Williams JM. Development and approval of new vaccines. Can Vet J 1990; 31:617-620.k 116. Shewen P, Sharp A, Wilkie BN. Efficacy testing a Pasteurella haemolytica extract vaccine. Vet Med 1988; 83:1078-1083.h 117. Sibbel RL, Bass EP, Thomas PC. How long will a killed IBR vaccine protect against challenge? Vet Med 1988; 83:90-92.^h 118. Simam PK, Kagumba M. The efficacy of lyophilized T1 vaccine against contagious bovine pleuropneumonia. Bull Anim Hlth Prod Afr 1989; 37:197-200.^{c,b,g,j} 119. Smith CK, Davidson JN, Henry CW. Evaluating a live vaccine for Pasteurella haemolytica in dairy calves. Vet Med 1985; 80:78-88.^{a,b,g} 120. Smith CK. The use of live cultures of Pasteurella haemolytica or Pasteurella multocida to immunize cattle against borespiratory disease. BovPractitioner 1988; 23:31-34.^h 121. Stewart RS, Gershwin LJ. Role of IgE in pathogenesis of bovine respiratory syncytial virus in sequential infection in vaccinated and nonvaccinated calves. Am J Vet Res 1989; 50:349-355.h,i 122. Stewart RS, Gershwin LJ. Systemic and secretory antibody responses to sequential bovine respiratory syncytial virus infections in vaccinated and nonvaccinated calves. Am J Vet Res 1990; 51:1596-1602.^{h,i} 123. Stockdale PHG, Langford EV, Darcel Cle Q. Experimental bovine pneumonic pasteurellosis I Prevention of the disease. Can J Comp Med 1979; 43:262-271.h 124. Stockdale PHG, Jericho KWF, Yates WDG, et al. Experimental bovine pneumonic pasteurellosis II Genesis and prevention. Can J Comp Med 1979;43:272-279.h 125. Stott EJ, Thomas LH, Taylor TG, et al. A comparison of three vaccines against respiratory syncytial virus in calves. J Hygiene 1984; 93:251-261.^h 126. Stott EJ, Thomas LH, Howard CJ, et al. Field trial of a quadrivalent vaccine against calf respiratory disease. Vet Record 1987;121:342-347. fg 127. Straub OC, Mawhinney IC. Vaccination to protect calves against infectious bovine rhinotracheitis. Vet Rec 1988; 122:407-411.h 128. Sutton ML. Rapid onset of immunity in cattle after intramuscular injection of a modified-live-virus. Vet Med Sm An Clin 1980; 75:1447-1456.^{a,b} 129. Syvrud B, Armstrong DA. Testing the efficacy of BRSV vaccination in herds with variable respiratory problems. Vet Med 1988; 83:429-430.^{a,b,c,e,f} 130. Talens LT, Beckenhauer WH, Thurber ET, et al. Efficacy of viral components of a nonabortigenic combination vaccine for prevention of respiratory and reproductive system diseases in cattle. J Am Vet Med Assn 1989;194:1273-1280.c.i 131. Thomas PC, Jones RH. Duration of immunity afforded by one dose of killed-virus bovine viral diarrhea vaccine. Agri-Practice 1985; 6:34-37.^{h,i} 132. Thomas PC, Peters E, Henning ER, et al. Evaluation of a modified live virus bovine respiratory syncytial virus vaccine used alone and in combination vaccines. Agri-Practice 1986; 7:26, 28-30. 133. Thomson JR, Nettleton PF, Greig A, et al. A bovine resvirus vaccination trial. Vet Record 1986;119:450-453.gj 134. Todd JD. Development of intranasal vaccination for the immunization of cattle against infectious bovine rhinotracheitis. Can Vet J 1974;15:257-259.* 135. Tyeryar FJ, Richardson LS, Belshe RB. Report of a workshop on respiratory syncytial virus and parainfluenza viruses. J Inf Dis 1978;137:835-846.k 136. Van Donkersgoed J, Potter AA, Mollison B, et al. The effect of a combined Pasteurella haemolytica and Haemophilus somnus vaccine and a modified-live bovine respiratory syncytial virus vaccine against enzootic pneumonia in young beef calves.Can Vet J 1994; 35:239-241.^j 137. Van Koevering MT, Gill DR, Owens FN, Smith RA, et al. Vaccine treatments to improve health and performance of newly arrived stocker cattle. Oklahoma State University Animal Science Research Report 1992; MP-136:342-346.^{a,b} 138. Verhoeff J, van Nieuwstadt APKMI. Prevention of bovine respiratory syncytial virus infection and clinical disease by vaccination. Vet Record 1984; 115:488-492.^{f,g} 139. Wilkie BN, Markham RJF, Shewen PE. Response of calves to lung challenge exposure with Pasteurella haemolytica after parenteral or pulmonary immunization. Am J Vet Res 1980; 41:1773-1778.h 140. Wilkie BN. Is immunization against bovine respiratory disease possible? Can Vet

zation of feedlot calves with a commercial Haemophilus somnus

resistance

respiratory

90.

J 1984;25:48-50.^k 141. Wilson SH. Why are meaningful field trials difficult to achieve for bovine respiratory disease vaccines? Can Vet J 1989; 30:299-302.^k 142. Woods GT, Sinha SK, McKeown JA, Brandly CA. Preshipment vaccination of feeder cattle with bovine para-influenza vaccine. JAm Vet Med Assn 1961; 139:1208-1211. a.b.c 143. Woods GT, Segre D, Barthel C, et al. The role of viral agents in respiratory diseases of cattle. II. A respiratory disease in feeder cattle vaccinated with Pasteurella bacterin an bovine para-influenza 3 vaccine before shipment. Am J Vet Res 1962; 23:987-991. a,b,cj 144. Woods GT, Mansfield ME, Segre D, et al. The role of viral agents in respiratory diseases of cattle. III. Respiratory disease in beef calves vaccinated before weaning with bovine myxovirus para-influenza 3 (SF-4) vaccine. Am J Vet Res 1962; 23:832-835. a.b.c 145. Woods GT, Mansfield ME, Cmarik G, et al. Vaccination of beef calves before weaning with bovine parainfluenza-3 (SF-4) vaccine in an adjuvant. Am J Vet Res 1964; 25:705-883.^{a,b,c,j} 146. Woods GT, Mansfield ME, Cmarik G, et al. Vaccination of beef calves before weaning with a live-virus bovine myxovirus parainfluenza-3 vaccine. Am J Vet Res 1968; 29:1349-1353.^{a,b,c,j} 147. Woods GT, Mansfield ME, Krone J. A controlled field study using live virus vaccines and an antiserum in a preconditioning program. Can J Comp Med 1972; 36:12-16.a,bj 148. Woods GT, Mansfield ME, Webb RJ. A three year comparison of acute respiratory disease, shrink and weight gain in preconditioned and non-preconditioned Illinois beef calves sold at the same auction and mixed in a feedlot. Can J Comp Med 1973: 37:249-255.a,e,m 149. Woods GT, Mansfield ME, Cmarik GF. Effect of certain biologic and antibacterial agents on development of acute respiratory tract disease in weaned beef calves. J Am Vet Med Assn 1973; 162:974-978.^b 150. Woods GT, Mansfield ME, Cmarik G, et al. Effects of bovine viral diarrhea and parainfluenza-3 virus vaccines on development of respiratory tract disease in calves. J Am Vet Med Assn 1973; 163:742-744.a,b,c 151. Woods GT, Mansfield ME, Krone J. Active and passive immunity to bovine viral respiratory diseases in beef calves after shipment. Can J Comp Med 1973; 37:336-340.bj 152. Woods GT, Mansfield ME, Cmarik GF, Marquis G. A four-year clinical and serologic study of the use of inactivated parainfluenza-3 virus vaccines and Pasteurella sp bacterins in beef calves. Vet Med Sm An Clinician 1974; 69:474-478. a,bj 153. Woods GT, Crandell RA, Mansfield ME. A comparison of immunologic response to intranasal and intramuscular parainfluenza-3 live virus vaccines in beef calves challenged experimentally in the feedlot. Res Comm Chemical Path Pharm 1975; 11:117-128, a,b,c,h,i 154. Woods GT.

Study of addition of live vaccines against bovine respiratory syncytial virus and Pasteurella hemolytica. Agri-Practice 1986; 7:11-16.b,^{ij} 155. Wright AJ, Mowat DN, et al. Supplemental chromium and bovine respiratory disease vaccines for stressed feeder calves. Can J An Sci 1994; 74:287-295.^j 156. Yates WDG, Stockdale PHG, Babiuk LA, et al. Prevention of experimental bovine pneumonic pasteurellosis with an extract of Pasteurella haemolytica. Can J Comp Med 1983; 47:250-256.h 157. York IA, Thorsen J. Evaluation of a subunit vaccine for bovine adenovirus type 3. Am J Vet Res 1992; 53:180-183.ⁱ 158. Zuffa A, Branyik A, Cernik K, et al. Protection against experimental infection of calves vaccinated intranasally by three attenuated strains of IBR virus. Zbl Vet B 1982; 29:413-425.h 159. Zygraich N, Vascoboinic E, Huygelen C. Immunity studies in calves vaccinated with a multivalent live respiratory vaccine composed of I.B.R., parainfluenza 3 and bovine adenovirus 3. Dev Biol Stand 1976; 33:379-383.1

- a. Blinding of assessors to treatment groups not mentioned.
- b. Method of assignment of experimental units to treatment groups not mentioned.
- c. No statistical analysis of data and insufficient information provided in paper to allow analysis.
- d. Inappropriate statistical test used to analyze data; appropriate test used incorrectly; P-values reported are incorrect.
- e. No control group or invalid control group.
- f. Inappropriate definition of experimental unit and pseudoreplication.
- g. Production setting and/or calf type and/or vaccine regime not practical or applicable to North American beef cattle production (external validity).
- h. Used experimental challenge model instead of field or simulated field exposure.
- i. Only reported outcomes such as antibody levels, seroconversion rates, immune function indicators, or product safety instead of clinically relevant outcomes, such as morbidity and mortality.
- $j. \ In adequate statistical power to detect significant differences if they existed.$
- k. No data presented, editorial, or review.
- Inadequate follow up of all animals that entered the field trial.
 m. Statistical methods not discussed or explained in materials and methods.