## Serum Protein Levels in Holstein Calves Fed Pasteurized-Frozen-Thawed or Unpasteurized First-milk Colostrum

Mario Medina-Cruz<sup>1</sup>, MVZ, MSc, DCV; C. Cruz<sup>3</sup>, MVZ; H. H. Montaldo<sup>2</sup>, MVZ, MC, PhD

<sup>1</sup>Departamento de Reproducción, Facultad de Medicina Veterinaria y Zootecnia, Universidad Nacional Autónoma de México. Circuito Exterior, Av. Universidad 3000, Col. Copilco, Del. Coyoacán, México DF 04505, MÉXICO Email: mmc@servidor.unam.mx, kingcheetah10@hotmail.com, Ph: 52(55)5622-5860 México (corresponding author) <sup>2</sup>Departamento de Genética y Estadística, Facultad de Medicina Veterinaria y Zootecnia, Universidad Nacional Autónoma de México. Circuito Exterior, Av. Universidad 3000, Col. Copilco, Del. Coyoacán, México DF 04505, MÉXICO.

<sup>3</sup>Private practice, Santiago de Querétaro, Qro, México

#### Abstract

This study compared serum protein (SP) concentrations in male and female Holstein calves fed pasteurized-frozen-thawed colostrum (PFTC) or unpasteurized colostrum (UPC) in a Mexican commercial dairy herd. To prepare PFTC, first-milk colostrum was collected and mixed to form 12 batches. All batches were pasteurized at 145°F (63°C) for 30 minutes and poured into 2.11 quarts (2 L) plastic bags for freezing for a minimum of 24 hours before use. Prior to feeding, frozen bags of colostrum were thawed in water at 122 to 131°F (50 to 55°C) for 15 minutes, and fed three times to each calf during the first 22.1 hours of life. Calves receiving UPC were hand-fed 1.74 quarts (1.65 L) of colostrum per feeding for three consecutive feedings with a suckling bottle in the first 22.9 hours of life. Blood from each calf was collected between 24 and 96 hours after birth. The SP level was  $5.92 \pm 0.15$  g/100 mL for calves receiving PFTC and  $6.53 \pm 0.14$  g/100 mL for those receiving UPC (P=0.0048). The percentage of calves with failure of passive transfer (FPT) of immunoglobulins with SP levels less than 5.2 g /100 mL was 24% for calves fed PFTC and 3.8% for calves fed UPC (P=0.06).

**Keywords:** bovine, calves, serum protein, colostrum, pasteurization, freezing

#### Résumé

Cette étude comparait la concentration sérique des protéines chez des veaux Holstein mâles et femelles nourris avec soit du colostrum pasteurisé congelé et puis dégelé ou soit du colostrum non-pasteurisé dans une ferme laitière commerciale mexicaine. Pour préparer le colostrum pasteurisé, le colostrum du premier lait a été recueilli et mélangé pour former 12 lots. Tous les lots ont été pasteurisés à 145°F (63°C) pendant 30 minutes

et versés dans des sacs en plastique de 2.11 pintes (2 L) pour congélation pendant au moins 24 heures avant leur utilisation. Avant l'abreuvement, les sacs congelés de colostrum ont été dégelés dans de l'eau entre 122 et 131°F (50 et 55°C) pendant 15 minutes et le colostrum a été donné trois fois à chaque veau pendant les premières 22.1 heures suivant la naissance. Les veaux qui recevaient le colostrum non-pasteurisé étaient nourris à la main avec 1.74 pintes (1.65 L) de colostrum par repas pendant trois repas consécutifs à l'aide d'un biberon pendant les premières 22.9 heures suivant la naissance. Un échantillon de sang a été prélevé de chaque veau entre 24 et 96 heures après la naissance. La concentration sérique des protéines était de 5.92 (±0.15) g/100 ml chez les veaux recevant le colostrum pasteurisé et de 6.53 (±0.14) g/100 ml chez ceux recevant le colostrum non-pasteurisé (P=0.0048). Le pourcentage de veaux pour lesquels le transfert passif n'a pas réussi pour les immunoglobulines avec des niveaux sériques de moins de 5.2 g/100 ml était de 24% chez les veaux recevant le colostrum pasteurisé et de 3.8% chez les veaux recevant le colostrum non-pasteurisé.

#### Introduction

Morbidity and mortality are much higher in calves with failure of passive transfer (FPT) when serum protein (SP) levels are less than 5.2 g/100 mL as compared to calves with SP levels above 5.5 g/100 mL.<sup>11,22,25</sup> The two main factors associated with higher SP levels are volume of colostrum fed and the total immunoglobulin mass in the colostrum.<sup>7</sup> First-milk colostrum has the highest immunoglobulin and protein concentrations compared to second and subsequent colostrum milkings.<sup>6,20</sup>

Specific pathogens can be transferred from cow to calf through colostrums, such as *Campylobacter* spp,<sup>13</sup> *Listeria monocytogenes*,<sup>3</sup> *Salmonella* spp,<sup>8</sup> *Mycobacterium avium* subsp paratuberculosis (MAP),<sup>21,23</sup> Mycobacterium

bovis,<sup>10,24</sup> other Mycoplasma spp, Staphylococcus spp, Streptococcus uberis and Escherichia coli.<sup>4</sup> Butler et al<sup>1</sup> demonstrated killing of Mycoplasma californicum, Mycoplasma bovis and Mycoplasma canadense by pasteurization of discarded milk at 149°F (65°C) for 60 minutes. Pasteurization also destroys a portion of the immunoglobulins present in colostrum. Meylan  $et al^{15}$  reported a mean loss of 12.3% of immunoglobulin content after heating small volumes of colostrum to 145°F (63°C) for 30 minutes. When the initial immunoglobulin levels are determined using a colostrometer, these authors suggested adjusting the volume of colostrum fed to calves to ensure successful passive transfer of immunity. The partial destruction of immunoglobulins during pasteurization in colostrum influences the SP levels in calves. Godden et al,<sup>9</sup> using on-farm commercial batch pasteurization of colostrum at 145°F (63°C) for 30 minutes and controlling for the time interval between feedings, reported significantly lower serum IgG concentrations in calves fed pasteurized colostrum (9.7 mg/mL) than for calves fed fresh colostrum (19.1 mg/mL). However, the use of hightemperature short-time (HTST) pasteurization at 161°F (72°C) for 15 seconds<sup>12</sup> resulted in similar mean serum IgG concentrations in treatment (1,476 mg/100 mL) and control (1,435 mg/100 mL) calves without differences in the proportion of calves with FPT between the treatment (16.22%) and control groups (19.55%).

Frequently, colostrum is frozen and thawed as a tool to feed high levels of immunoglobulins on many dairies without evidence of detrimental effects upon immunoglobulins or nutrients.<sup>2,17</sup> According to Foley et al,<sup>5</sup> there was no loss of vitamin A and only a small reduction in carotene content (6%) in colostrum stored at  $-5^{\circ}$ F (-20.5°C) for six months. When Holloway et al<sup>11</sup> fed calves at three hours of age with 1.05 gallon (4 L) aliquots of frozen-thawed or fresh colostrum using oroesophageal intubations, no significant differences in serum IgG concentrations were found at 48 hours of age  $(2,097.0 \pm 681.7 \text{ mg}/100 \text{ mL vs } 1,734.3 \pm 958.6 \text{ mg}/100$ mL, respectively). When feeding the dam's colostrum is not possible, strategies to prevent transmission of infectious agents through the colostrum include feeding pasteurized colostrum or colostrum replacers.<sup>9</sup> To our knowledge there are no studies of feeding pasteurizedfrozen-thawed colostrum (PFTC) to calves to evaluate the effect upon SP levels.

The objective of this study was to determine and compare the SP concentrations in calves fed either PFTC or UPC.

#### **Materials and Methods**

#### Colostrum Collection

This study was conducted on a commercial dairy in the state of Queretaro, Mexico. The milking herd con-

sisted of 1,250 cows, and cows calved on a farm located three miles from the milking herd. Calvings occurred in individual maternity pens located within the closeup corrals. All cows that calved during the night were milked-out by machine and colostrums were kept and stored in individually identified buckets at room temperature (41-50°F; 5-10°C) before transport to the dairy herd, where pasteurization was carried out the next morning. As a general rule, colostrums from overnight calvings were pasteurized and colostrums from daytime calvings were fed fresh to the calves. Three days after calving, the cows were transported to the main dairy and placed in the milking herd.

#### Management of Pasteurized Frozen-thawed Colostrums (PFTC) and Unpasteurized Colostrum (UPC)

First-milk colostrum obtained from each cow calving during nighttime was visually inspected and smelled prior to pasteurization. Bloody, mastitic or soured colostrum was discarded. For all viable colostrums, a colostrometry test<sup>a</sup> was performed at a range in temperature of 68-75°F (20-24°C), and only colostrums with high specific gravity  $(\geq 1.054)$  and high immunoglobulin contents  $(\geq 70 \text{ g/L})$  were included in the study. These colostrums were filtered through a mesh and mixed to form a batch per day, for a total of 12 batches during the experiment. For every batch, volume was recorded and the colostrometry test was performed again at a temperature of 68-75°F (20-24°C). Pasteurization was carried out using an automated commercial batch-type pasteurizer<sup>b</sup> that heated the contents to 145°F (63°C) for 30 minutes, and then cooled to 39.2°F (4°C). Once pasteurization was completed, consistency of the colostrum was assessed as normal, slightly thickened or moderately thickened, and then was poured into 2.11 quart (2 L), zip-lock type plastic bags identified with the date and batch number. Bags were immediately placed in a freezer at 20°F (-7°C). The following day, the frozen colostrums were transported to the calf farm where they were stored in another freezer at -4°F (-20°C). These colostrums were kept frozen for a minimum of 24 hours before thawing to feed calves.

Cows calving during the day were immediately milked out (first-milk colostrums) and colostrum were filtered through an extended gauze. This UPC was not pooled, but instead colostrum from each cow was fed to her own calf without performing the colostrometry test.

#### Calf Allocation and Treatment, Colostrum Thawing, Sample Collection, Tests and Record Keeping

All calves (40 males and 11 females) born from October 1 to October 30, 2004 and included in the study were immediately placed in clean, individual hutches after birth. Calves were enrolled alternatively according to the order of birth into either the group receiving PFTC or the control group receiving UPC. Bags containing 2.11 quarts (2 L) of PFTC were thawed by immersion in warm water at 122-131°F (50-55°C) for 10 to 15 minutes. Once close to or at body temperature, 2.11 quarts of PFTC was fed to calves in the experimental group, using a suckling bottle, for three consecutive feedings during the first 24 hours of life. Control calves were fed 1.74 quarts (1.65 L) of UPC using a suckling bottle for three consecutive feedings during the first 24 hours of life.

Between 24 and 96 hours after birth, blood was obtained from every calf by jugular venipuncture for serum collection and a handheld refractometer<sup>c</sup> with temperature compensation was used to determine SP<sup>14</sup> values.

Information recorded for every calf included calf identification number, date and time of birth, gender, treatment allocation, volume fed in the first, second and third feedings and time of each feeding. Age (hours) and total volume fed in the first 24 hours of life were calculated.

From October 1 to 15, 2004, colostrum pasteurization, freezing, blood sample collection and record keeping were done by the first author, and from October 16 to 30, 2004 by the second author. Throughout the study at least one of the authors provided daily supervision of colostrum collection, calf allocation, colostrum thawing and feeding, and record keeping at the farm.

#### Statistical Analysis

Of the 51 calves born during the study period, 25 calves were fed PFTC and 26 were fed UPC. The statistical analysis was performed with linear models (ANOVA) using the General Model Procedure.<sup>19</sup> The analysis examined the associations between colostrum assignment and SP levels. Final model for analysis of the dependent variable SP (g/100 mL) included only the treatment effect. The effects of time to first feeding (in hours), calf gender, calf age at sampling (hours) as linear and quadratic covariates and all first-order interactions were excluded from the final model because they were not found significant (P>0.05).

#### Results

#### Colostrum

The average volume of the 12 batches of first-milk colostrum was  $5.7 \pm 2.2$  gallons (21.6  $\pm$  8.5 L), and average immunoglobulin content was  $85.2 \pm 18.9$  g/L. Consistency after pasteurization remained normal in six samples, slightly thickened in four and moderately thickened in two. Colostrum ingestion occurred at 1.3, 12.2 and 22.1 hours of life in calves fed PFTC, and at 1.1, 12.4 and 22.9 hours for calves fed UPC. There was no significant difference in colostrum ingestion time between treatment groups (P>0.05). Average volume consumed per feeding in the first day of life was 2.11 quarts (2 L) for calves fed PFTC and 1.74 quarts (1.65 L) for those fed UPC, for a total of 1.58 gallons (6 L) and 1.30 gallons (4.95 L), respectively.

#### Serum Protein

Average SP level was lower in calves consuming PFTC ( $5.92 \pm 0.15 \text{ g}/100 \text{ mL}$ ) compared to those fed UPC ( $6.53 \pm 0.14 \text{ g}/100 \text{ mL}$ ), as shown in Table 1 (P=0.0048). The percentage of calves with FPT (SP level <5.2 g/100 mL) tended to be higher for the calves fed PFTC (24%) compared to calves fed UPC (3.8%) (Table 2; P=0.06).

#### Discussion

At the beginning of this study, three batches of pooled colostrums were discarded due to extreme thickening after pasteurization. In the 12 batches included in the study, there was moderate thickening in two batches, which resulted in some difficulty in consumption by the calves, although they consumed all that was offered. The first batch included in the study had a volume of 11.88 gallons (45 L) and an immunoglobulin content of 42 g/L. However, if we separate these data, the average volume for the remaining 11 batches was of  $5.0 \pm 1.3$  gallons (19.7  $\pm 5$  L), with a minimum of 2.38 and a maximum of 7.4 gallons (9-28 L). The average immunoglobulin content was of  $88.8 \pm 14.5$  g/L, with a minimum of 70 g/L and a

**Table 1.** Serum protein levels in calves fed pasteurized-frozen-thawed colostrum or unpasteurized colostrum for the first three feedings within the first 24 hours of life.

Group	Calves Serum protein (g		ein (g/100 mL)
	n	X	Std error
Pasteurized-frozen-thawed colostrum	25	5.92 ª	0.15
Unpasteurized colostrum	26	6.53 <sup>b</sup>	0.14

<sup>ab</sup> Different superscripts indicate statistically significant differences (P=0.0048)

	$\bigcirc$
	© Coj
5	pyrig
%	;ht A
68	mei
92.4	102
	ın /
	Ass
	soc
	iat
fed PFTC	ion
s and had	of
lins. The	·B
re higher	ovi
suggested	ne
nt passive	Pr
asteuriza-	act
l thawing.	itic
stroy a sig-	one
strums, as	rs;
igley. <sup>18</sup> In	do
adequate	en
din $et al^{12}$	ac
5 seconds,	ces
centage of	SS (
measured	list
een treat-	rib
our study f in calves	uti
for calves	on
tor carves	•

Table 2. Proportion of calves fed pasteurized frozen-thawed colostrums that had serum protein	n levels of <5.2, 5.2-
5.5 or >5.5 g/100 mL.*	

Calves Serum protein (g/100 mL) Group 5.2-5.5 < 5.2 > 5.5 % % n n n n 25 6 24 2 8 17 Pasteurized-frozen-thawed colostrum 3.8 1 3.8 Unpasteurized colostrum 26 1 24

\* Independence chi-square test, P=0.06

maximum 110 g/L. The sensitivity of the colostrometry test increases in colostrums with higher specific gravity.<sup>16</sup> In this study, and based only on the colostrometry test, we purposely selected only colostrums with a specific gravity equal or greater than 1.054. This resulted in a high immunoglobulin content (>70 g/L), with the exception of the first batch.

The use of small-sized batches in our study was due to either a low number of calvings per night, insufficient production of colostrums of high specific gravity or discarded colostrums due to blood or mastitis. According to Godden *et al*,<sup>9</sup> smaller-volume batches are associated with less loss in immunoglobulin content in pasteurized colostrums.

The differences in hours at first, second and third feedings between the two treatment groups were not significant (P>0.05). In the group fed PFTC, three plastic bags containing 2.11 quarts of frozen colostrum were thawed and fed during the first day of life to each calf. Calves fed UPC received colostrums from their own dams. Calves in the UPC group were fed 1.74 quarts (1.65 L) three times in the first day of life, resulting in a total volume of 1.30 gallons (4.95 L). In spite of the smaller volume of UPC fed to the control calves, the SP levels attained by this group were higher than those in calves receiving PFTC, suggesting that the immunoglobulin concentrations of the UPC were higher than those in PFTC.

Godden et al,<sup>9</sup> found significantly higher IgG concentrations in calves fed unpasteurized colostrum (19.1 mg/mL) than in those fed pasteurized (9.7 mg/mL) colostrum. In our study, calves fed UPC were handled and fed according to the farm's management protocol, which explains why the colostrum given to these animals was not evaluated with a colostrometer. In spite of this, calves that received UPC from their own dams had higher SP (P=0.0048) levels (6.53 ± 0.14 g/100 mL) than calves fed PFTC (5.92 ± 0.15 g/100 mL). Results from this study suggest that calves fed at least 1.30 gallons (4.95 L) of first-milk UPC within the first 23 hours of

life achieved a satisfactory SP level. Calves fed PFTC also developed satisfactory SP concentrations and had successful passive transfer of immunoglobulins. The levels of SP in both groups in this study are higher than those recommended by Tyler *et al*,<sup>22</sup> who suggested that 5.5 g/100 mL are indicative of an efficient passive transfer in the calf.

PFTC was subjected to three processes: pa tion, freezing for a minimum of 24 hours and The two latter processes apparently did not des nificant level of immunoglobulins in the colos has been suggested by Holloway et al<sup>11</sup> and Qu our study, frozen-thawed colostrums were an source of IgG for Holstein calves. Jamaludo pasteurized colostrum at 161°F (72°C) for 18 and reported no significant differences in percalves experiencing FPT (<10 mg/mL of IgG measured 48 to 96 hours after colostrum intake) between treatment (16.2%) and control groups (19.5%). In our study we found a higher percentage (P=0.06) of FPT in calves fed PFTC; 24% for calves fed PFTC and 3.8% for calves fed UPC. Differences reported in studies may be due to variations in pasteurization protocols.

Finally, freezing colostrum requires a freezer, extra handling, daily thawing of required colostrum and is less cost-effective than refrigeration.<sup>2,5,18</sup> Use of PFTC, however, can provide a source of immunoglobulins when fresh colostrum is in short supply.

#### Conclusions

We acknowledge some aspects of our study might limit the value of the results, such as the use of a colostrometer for ruling out colostrums with low immunoglobulins content, differences in handling the colostrums between cows calving during night hours and in daylight, feeding a smaller volume to calves fed UPC and variation in batch sizes of PFTC which were determined by the number of suitable colostrums available from the night before. The selection of first-milk colostrums of high immunoglobulin content for pasteurization at  $145^{\circ}F(63^{\circ}C)$ for 30 minutes, followed by freezing, thawing and then feeding to just-born calves, provided SP levels 0.42 g/100mL or 10.3% higher than the minimum levels recommended to avoid FPT. These levels, however, were lower than those of the calves fed UPC. Likewise, a strong trend for a higher percentage of calves with FPT was found in the group fed PFTC, but the difference was not significant (P=0.06).

#### Endnotes

<sup>a</sup>Colostrometer, Biogenics, Mapleton, OR 97453

<sup>b</sup>DT Silver pasteurizer, Dairy Tech. Inc. Windsor, CO 80550

<sup>e</sup>Refractometer, TS 400, PO Box 123, Buffalo, NY 14240

#### Acknowledgements

Financing for acquisition of the pasteurizer and other materials and equipment used was made possible thanks to funding provided by PAPIIT, DGAPA and UNAM through research project IN: 218701.

#### References

1. Butler JA, Sickles SA, Johanns CJ, Rosenbusch RF: Pasteurization of discard mycoplasma mastitic milk used to feed calves: thermal effects on various mycoplasma. *J Dairy Sci* 83:2285-2288, 2000.

2. Carlson SMA, Muller LD: Compositional and metabolic evaluation of colostrum preserved by four methods during warm ambient temperatures. J Dairy Sci 60:566-571, 1996.

3. Farber JM, Sanders GW, Malcolm SA: The presence of *Listeria* spp in raw milk in Ontario. *Can J Microbiol* 34:95-100, 1988.

4. Fecteau G, Baillargeon P, Higgins R, Paré J, Fortin M: Bacterial contamination of colostrum fed to newborn calves in Québec dairy herds. *Can Vet J* 43:523-527, 2002.

5. Foley JA, Otterby DE: Availability, storage, treatment, composition and feeding value of surplus colostrum: a review. *J Dairy Sci* 61:1033-1060, 1978.

6. Gay CC, Besser TE, Pritchett LC, Hancock DD, Wikse S: Avoidance of passive transfer failure in calves. *Proc Am Assoc Bov Pract* 20:118-120, 1988.

7. Gay CC: Failure of passive transfer of colostral immunoglobulins and neonatal disease in calves: a review. *Proc Fourth International Symp Neonatal Diarrhea*. Veterinary Infectious Disease Organization. pp 346-364, 1983. 8. Giles N, Hopper SA, Wray C: Persistence of *S. typhimurium* in a large dairy herd. *Epidemiol Infect* 103:235-241, 1989.

9. Godden SM, Smith S, Feirtag JM, Green LR, Wells SJ, Fetrow JP: Effect of on-farm commercial batch pasteurization of colostrum on colostrum and serum immunoglobulin concentration in dairy calves. J Dairy Sci 86:1503-1512, 2003.

10. Grant IR, Ball HJ, Rowe MT: Thermal inactivation of several *Mycobacterium* spp in milk by pasteurization. *Appl Microbiol* 22:253-256, 1996.

11. Holloway NM, Tyler JW, Lakritz J, Carlson SL, Holle J: Serum immunoglobulin G concentrations in calves fed fresh and frozen colostrum. J Am Vet Med Assoc 219:357-359, 2001.

12. Jamaluddin AA: Effects of feeding pasteurized colostrum and pasteurized waste milk on mortality, morbidity, and weight gain of dairy calves: field trial and economic analysis. Ph.D. Diss. University of California, Davis, USA, 1995.

13. Lovett J, Francis DW, Hunt JM: Isolation of *Campylobacter jejuni* from raw milk. *Appl Environ Microbiol* 46:459-462, 1983.

14. McBeath DG, Penhale WJ, Logan EF: An examination of the influence of husbandry on the plasma immunoglobulin level of the newborn calf, using a rapid refractometer test for assessing immunoglobulin content. *Vet Rec* 88:266-270, 1971.

15. Meylan M, Rings M, Shulaw WP, Kowalski JJ, Bech-Nielsen S, Hoffsis GF: Survival of *Mycobacterium paratuberculosis* and preservation of immunoglobulin G in bovine colostrum under experimental conditions simulating pasteurization. *Am J Vet Res* 57:1580-1585, 1995.

16. Pritchett LC, Gay CC, Hancock DD, Besser TE: Evaluation of the hydrometer for testing immunoglobulin G1 concentrations in Holstein colostrum. *J Dairy Sci* 77:1761-1767, 1994.

17. Quigley J: http://www.calfnotes.com/ Calf Note # 96, Pasteurized colostrum, 2003.

18. Quigley J: Calving ease. http://www.calfnotes.com/CNcalvingease. htm Pro's and Con's of Feeding Frozen Colostrum, July 2004.

19. SAS Institute: SAS User's Guide: Statistics, Version 8 ed. SAS Inst, Inc, Cary, NC, 1999.

20. Stott GH, Fleenor WA, Kleese WC: Colostral immunoglobulin concentration in two fractions of first milking postpartum and five additional milkings. *J Dairy Sci* 64:459-465, 1981.

21. Streeter RN, Hoffsis GF, Bech-Nielsen S, Shulaw WP, Rings DM: Isolation of *Mycobacterium paratuberculosis* from colostrum and milk of subclinically infected cows. *Am J Vet Res* 56:1322-1324, 1995.

22. Tyler JW, Hancock DD, Parish SM, *et al*: Evaluation of 3 assays for failure of passive transfer in calves. *J Vet Int Med* 10(5):304-307, 1996.

23. Tyler JW, Lakritz J, Hostetler DE, *et al*: Effect of pasteurization at 76 and 63°C on the absorption of colostral IgG in calves. *J Dairy Res* 67:619-423, 2000.

24. Walz PH, Mullaney TP, Render JA, Walker RD, Mosser T, Baker JC: Otitis media in preweaned Holstein dairy calves in Michigan due to *Mycoplasma bovis*. J Vet Diagn Invest 9:250-254, 1997.

25. Weaver DM, Tyler DC, VanMetre DC, Hostetler DE, Barrington GM: Passive transfer of colostral immunoglobulins in calves. J Vet Intern Med 14:569-577, 2000.

ISSN 0524-1685



### Guidelines for Authors

Two issues of *The Bovine Practitioner* are published annually, one in the spring and one in the summer. It also serves as a communication medium between bovine practitioner organizations around the world. All manuscripts and communications must be presented in English.

Most articles in the journal are peer-reviewed or refereed. Papers submitted for publication in the peer-reviewed section are anonymously reviewed by three members of the editorial board. In some cases, papers may be reviewed by an outside expert(s) who is not a regular member of the editorial board. Papers published in the peer-reviewed section of the journal will be identified with a "Peer-Reviewed" banner at the top of the first page. Papers rejected by the editorial board for publication as peer-reviewed articles do not automatically qualify for publication in the non-peer-reviewed sections.

Articles published in *The Bovine Practitioner* are intended to address the needs of bovine practitioners. Types of articles considered appropriate for the journal include research reports, case reports, review articles, retrospective studies and articles describing new techniques.

All papers should begin with an abstract. Research reports should follow with an introduction, materials and methods (including experimental design and statistical analysis), results, discussion and conclusions. At the author's discretion, results and discussion may be combined.

Case reports should be written to include an introduction, history, clinical findings, appropriate laboratory data, surgical/therapeutic management, discussion and conclusions.

Review articles covering topics important to the practitioner are welcome. They should address more recent advances and bring the reader cutting edge information related to bovine practice or to beef or dairy production.

Papers reporting retrospective studies should include an introduction, clinical implications or objectives of the study, the methodology used to evaluate the data, a section that details the significance of the findings to the practitioner and conclusions.

Two manuscripts and a diskette or CD should be submitted to the editor through the mail or via a parcel delivery service. Manuscripts should be double-spaced, using 12-point Times type and 1-inch margins. Both lines and pages should be numbered. When possible Microsoft Word should be used.

Figures, tables and photographs are welcome. Figures should be numbered on the back: legends for figures should be submitted on a separate sheet of paper. Photographs can be submitted as digital images or prints; prints are preferred over 2x2 slides.

English units of measure should be used for weights, measures and temperature. If the author desires, it is acceptable to follow English units with metric units in parenthesis, i.e....440 lb (200 kg) steer had a rectal temperature of 101.5°F (38.6°C). When the use of brand names is necessary, they should be listed in footnotes or endnotes, including the name of product, manufacturer, and manufacturer's city and state.

References to literature cited in the paper must be identified in the text by the use of superscripts. References should be listed in **alphabetical order**. Suggested style for citations in the reference section is as follows:

 Allen WM, Sansom BF: Parturient paresis (milk fever) and hypocalcemia (cows, ewes, and goats), in Howard JL (ed): Current Veterinary Therapy III. Food Animal Practice. Philadelphia, WB Saunders Co, 1993, pp 304-308.
Barth AD, Cates WF, Harland RJ: The effect of body fat and loss of fat on breeding soundness classification of beef bulls. Can Vet J 36:758-764, 1995.

3. Nutrient Requirements of Beef Cattle, ed 7. Washington DC, National Academy Press, 1996.

4. Syvrud R: Vaccination for bovine respiratory syncytial virus: Benefits for both cow/calf and feedlot cattle. *Proc* Am Assoc Bov Pract 21:204-206, 1989.

All correspondence and manuscripts should be addressed to:

The Bovine Practitioner Dr. Bob Smith, Editor 3404 Live Oak Lane Stillwater, OK 74075 405-372-8666 Office 405-743-8422 Fax

Rev 04/07

# THE AT COM SERVICE



# DISCOVER AN EXTRA CALF IN EVERY BOTTLE

Vaccinating with Vision® 7 results in 14 pounds more weaning weight per calf than Ultrabac® 7 — Your clients get an extra calf in every 50-dose bottle.

Blackleg vaccination stress can decrease performance, but Vision<sup>®</sup> minimizes the negative impact of vaccination.

The proof is in the pounds. Studies show:

- → Less stress in calves vaccinated with Vision 2mL compared to 5mL blackleg vaccines results in a weaning weight advantage from 9 to 18 pounds
- → Vision 7 2mL resulted in a 14-pound weaning weight advantage when compared to Ultrabac 7 5mL\* – that's an extra calf in every 50-dose bottle

Help your clients take the stress out of blackleg protection. Switch your recommendation to Vision vaccine today.

\* Intervet Technical Reference 93-9: Weaning Weight Comparison of Vision 7 and Ultrabac 7 in a Wyoming Beef Herd



P.O. Box 318 29160 Intervet Lane Millsboro, Delaware 19966-0318 intervetusa.com 800.441.8272 Vision is a registered trademark of Intervet Inc. or an affiliate. Ultrabac is a registered trademark of Pfizer Inc. ©2008 Intervet Inc. All rights.reserved. 1/08 0&B Part #32990V





## Does your fresh cow program cover everything it should?

We can help you put together a program that does. It starts with a complete protocol developed to help identify and address issues before they become costly problems. So cows stay in the milking string, not the sick pen. As part of the Pfizer Dairy Wellness Plan, we support fresh cow programs with products like EXCEDE® (ceftiofur crystalline free acid) Sterile Suspension for extended BRD therapy with just a single dose, and EXCENEL® RTU EZ (ceftiofur hydrochloride) Sterile Suspension for metritis, which is now significantly more syringeable! See your veterinarian to evaluate your own fresh cow program and visit dairywellnessplan.com to learn more.





As with all drugs, the use of EXCEDE Sterile Suspension is contraindicated in animals previously found to be hypersensitive to the drug. Though safe in cattle when properly administered, inadvertent intra-arterial injection in the ear is possible and is fatal. EXCEDE has a pre-slaughter withdrawal time of 13 days. As with all drugs, EXCENEL RTU EZ should not be used in animals found to be hypersensitive to the product. EXCENEL RTU EZ has a pre-slaughter withdrawal period of 3 days.

1 Based on the results of a blinded well-controlled study. Data on file, Study 0788-7922-2003-001, Pfizer Inc.

Dairy Wellness Makes a Difference is a trademark of Pfizer Inc. EXCEDE and EXCENEL are registered trademarks of Pharmacia and Upjohn Company LLC, a division of Pfizer Inc. ©2008 Pfizer Inc. All rights reserved. EXD08018

Full product information can be found on page 197